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(54) Title: COMPOSITIONS AND METHODS FOR PRODUCING ANTIGEN-PRESENTING CELLS

(57) Abstract: The present invention relates to compositions and methods for producing antigen-presenting cells, in vitro, ex vivo or in vivo. This invention relates more particularly to methods and compositions for producing dendritic cells using interleukin-15, preferably in combination with a growth factor such as Granulocyte-Macrophage colony stimulating factor. This invention is particularly suited for producing immature dendritic cells and activated cells from precursors, in vitro, ex vivo or in vivo. The invention also relates to compositions for implementing these methods, as well as compositions comprising antigen-presenting cells and uses thereof. Dendritic cells or membrane vesicles derived therefrom have utility in many applications, including diagnostic, therapy, vaccination, research, screening and gene delivery.

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COMPOSITIONS AND METHODS FOR PRODUCING ANTIGEN-PRESENTING CELLS

CROSS-REFERENCE TO RELATED APPLICATION

This application claims the benefit of U.S. Provisional Patent Application No.
5 60/203,571, filed May 11, 2000.

TECHNICAL FIELD OF INVENTION

The present invention relates to compositions and methods for producing antigen-presenting cells, in vitro, ex vivo or in vivo. This invention relates more particularly to methods and compositions for producing antigen-presenting cells using interleukin-15
10 (IL-15), preferably in combination with a growth factor such as Granulocyte-Macrophage Colony Stimulating Factor (GM-CSF). This invention is particularly suited for producing dendritic cells from precursors in vitro, ex vivo or in vivo. The invention also relates to compositions for implementing these methods, as well as compositions comprising antigen-presenting cells and uses thereof.

15 BACKGROUND OF THE INVENTION

The immune system comprises various cell populations, including antigen-presenting cells which are involved in antigen presentation to effector or regulatory cells, such as macrophages, B-lymphocytes and dendritic cells. Dendritic cells are potent antigen-presenting cells, involved in innate immunity, and capable of inducing antigen-
20 specific cytotoxic T lymphocyte (CTL) responses. For this reason, dendritic cells are being proposed for use in immunotherapy of various pathologies, including tumors or immune diseases, through production of said cells in vitro, sensitization to an antigen, and re-infusion to a patient. Also, membrane vesicles derived from dendritic cells have been discovered and isolated, which exhibit very powerful immunogenic activity (International
25 Application No. WO99/03499). Current dendritic cell production methods essentially rely on the differentiation from precursor cells (essentially peripheral blood monocytes) by culture in the presence a combination of a cytokine and a growth factor, more specifically interleukin-4 or interleukin-13 in combination with Granulocyte-Macrophage Colony

Stimulating Factor (GM-CSF). There is a need in the art to develop alternative production methods, which produce functional dendritic cells or functional antigen-presenting cells for use in any therapeutic, diagnostic, prophylactic or research area.

SUMMARY OF INVENTION

5 In one aspect, the present invention is a method for producing dendritic cells from dendritic cell precursors in vitro or ex vivo comprising contacting dendritic cell precursors with a composition comprising interleukin-15 and at least one growth factor. In one embodiment, the resulting dendritic cells are immature. These immature dendritic cells can be sensitized to an antigen. These immature dendritic cells can be administered to a
10 patient for modulating an immune response in the patient. The invention also includes a composition comprising immature dendritic cells produced according to this method.

In another aspect, the invention is a method for producing activated dendritic cells from dendritic cell precursors in vitro or ex vivo comprising contacting dendritic cell precursors with a composition comprising interleukin-15 and at least one growth factor to
15 form activated dendritic cells. The activated dendritic cells can be sensitized to an antigen. These activated dendritic cells can be administered to a patient for modulating an immune response in the patient. The invention also includes a composition comprising activated dendritic cells produced according to this method.

In another aspect, the invention is a method for producing mature dendritic cells
20 from dendritic cell precursors in vitro or ex vivo comprising contacting dendritic cell precursors with a composition comprising interleukin-15 and at least one growth factor to form immature dendritic cells and culturing the immature dendritic cells in the presence of an maturation agent and under conditions to promote maturation of the immature dendritic cells to form mature dendritic cells. In one embodiment, the immature dendritic cells are
25 sensitized to an antigen either prior to, during or after maturation. Exemplary maturation agents for in vitro use include lipopolysaccharide, CD40L, and poly(I):(C). These mature dendritic cells can be administered to a patient for modulating an immune response in the patient. The invention also includes a composition comprising mature dendritic cells produced according to this method.

In another aspect, the invention is a method for producing isolated membrane vesicles comprising contacting dendritic cell precursors with a composition comprising interleukin-15 and at least one growth factor to form immature dendritic cells, culturing the immature dendritic cells in the presence of at least one stimulation factor to promote
5 the release of membrane vesicles; and isolating the membrane vesicles. In one embodiment, the immature dendritic cells are sensitized to an antigen prior to or after membrane vesicle release. Exemplary stimulation factors include a suitable cytokine, irradiation, and low pH. These membrane vesicles can be administered to a patient for modulating an immune response in the patient. The invention also includes a composition
10 comprising membrane vesicles produced according to this method.

In another aspect, the invention is a method for producing immature dendritic cells from dendritic cell precursors in a patient comprising concurrent or sequential administration of interleukin-15 and at least one growth factor to the patient.

In another aspect, the invention is a method for producing activated dendritic cells
15 from dendritic cell precursors in a patient comprising concurrent or sequential administration of interleukin-15 and at least one growth factor to the patient.

In another aspect, the invention is a method for producing mature dendritic cells from dendritic cell precursors in a patient comprising concurrent or sequential administration of interleukin-15, at least one growth factor, and at least one maturation
20 factor to the patient.

In another aspect, the invention is a method for modulating an immune response in a patient via dendritic cells comprising concurrent or sequential administration of interleukin-15 and at least one growth factor. This method can also include administering at least one maturation factor to the patient.

25 In another aspect, the invention is a method for inducing or stimulating dendritic cell differentiation from precursors thereof comprising contacting the precursors with interleukin-15 and at least one growth factor.

In another aspect, the invention is a method for producing antigen-specific cytotoxic T lymphocytes in vitro or ex vivo comprising providing antigen-sensitized

dendritic cells prepared by contacting precursors thereof with interleukin-15 and at least one growth factor and subsequent sensitizing of the dendritic cells to an antigen, and contacting the antigen-sensitized dendritic cells with a population of cells comprising T lymphocytes to form activated antigen-specific cytotoxic T lymphocytes. These antigen-specific cytotoxic T lymphocytes can be administered to a patient for modulating an immune response in the patient. The invention also includes a composition comprising antigen-specific cytotoxic T lymphocytes produced according to this method.

In another aspect, the invention is a method for producing antigen-specific CD4+ T lymphocytes in vitro or ex vivo comprising providing antigen-sensitized dendritic cells prepared by contacting precursors thereof with interleukin-15 and a growth factor and subsequent sensitizing of the dendritic cells to an antigen and contacting the antigen-sensitized dendritic cells with a population of cells comprising T lymphocytes to form activated antigen-specific CD4+ T lymphocytes. These antigen-specific CD4+ T lymphocytes can be administered to a patient for modulating an immune response in the patient. The invention also includes a composition comprising antigen-specific CD4+ T lymphocytes produced according to this method.

In another aspect, the invention is a method for producing dendritic cell precursors comprising contacting monocytes in vitro or ex vivo with interleukin-15 to form dendritic cell precursors. The invention also includes a composition comprising dendritic cell precursors produced according to this method.

The interleukin-15 useful in the methods and compositions of the present invention is preferably selected from the group consisting of isolated interleukin-15, a recombinant interleukin-15 polypeptide, a nucleic acid, either wild type or engineered, encoding interleukin-15 in a suitable expression vector, and a cell population expressing a nucleic acid encoding interleukin-15. The growth factor useful in the methods and compositions of the present invention is preferably selected from the group consisting of isolated growth factor, a recombinant growth factor polypeptide, a nucleic acid encoding a growth factor in a suitable expression vector, and a cell population expressing a nucleic acid encoding a growth factor. More preferably, the growth factor is granulocyte-macrophage colony

stimulating factor. The maturation factor useful in the present invention is preferably selected from the group consisting of lipopolysaccharide, CD40L, and poly(I):(C).

BRIEF DESCRIPTION OF THE DRAWINGS

Fig. 1A and 1B depict the development of DCs in the presence of GM-CSF and IL-15 (GM-CSF+IL-15). Enriched CD14⁺ cells were cultured in the presence of GM-CSF and IL-15 over six days. Fig. 1A depicts non-adherent cells harvested and analyzed for CD1a, CD14, and HLA-DR by FACS Calibur (Becton Dickinson, San Jose, CA, US). Fig. 1B depicts cultured monocytes in GM-CSF and IL-15 analyzed in a time kinetic manner by FACS Calibur.

Fig. 2A and 2B depict maturation of GM-CSF+IL-15-derived DCs with lipopolysaccharide (LPS). Fig. 2A depicts the results obtained on Day 5 when a part of GM-CSF+IL-15-derived DCs were activated with LPS (100 ng/ml) and analyzed for HLA-ABC, CD40, CD80, CD83, and CD86. Fig. 2B depicts the results of FITC-Dextran up-take studies. Activated (dotted line) or non activated DCs (solid line) were incubated with 100 µg/ml FITC-Dextran for 30 min at 37°C. Control DCs were incubated on ice for 30 min as well. Afterwards, DCs were washed three times with PBS and analyzed for FITC-Dextran up-take by FACS Calibur.

Fig. 3A depicts partial expression of Langerin (marker of Langerhans cells, a special type of dendritic cells) and CCR6 (chemokine receptor 6) in GM-CSF+IL-15-derived DCs. Fig. 3B depicts GM-CSF+IL-15-induced DCs expressing higher levels of Langerin when compared to GM-CFS+IL-4/TGFβ-induced DCs. Representative of three experiments, Fig. 3C depicts the fact that the generation of Langerhans cell-like DCs with GM-CSF and IL-15 is independent of TGFβ-1. Monocytes were cultured for 6 days with cytokines shown above the histograms with 300 ng/ml daily or without anti-TGFβ-1 antibody and stained as indicated. Fig. 3D depicts the fact that GM-CSF+IL-15-induced cells, but not GM-CSF+IL-4-induced cells, have Langerhans cells phenotype.

Fig. 4A, 4B, and 4C depict the allostimulatory capacity of GM-CSF+IL-15-derived DCs. DCs activated with poly (I):(C) or non-activated DCs were cultured as described herein and co-cultured with allogeneic CD4⁺ (Fig. 4A) or CD8⁺ T cells (1 x 10⁵)

for 5 days (Fig. 4B). Cultures were pulsed with thymidine for 16 hours. Fig. 4C depicts a comparison of DCs cultured in GM-CSF+IL-15, GM-CSF+IL4, GM-CSF+IL-4+TGF β -1, or GM-CSF alone in an allogeneic MLR.

Fig. 5A and 5B depict presentation of TT (tetanus toxoid) or dying cell bodies by
5 GM-CSF+IL-15-derived DCs. DCs were cultured as described herein. GM-CSF+IL-15-derived DCs and GM-CSF+IL-4-derived DCs were treated with TT (Fig. 5A) or dying cell bodies derived from Me 275 (Fig. 5B) at 37°C for 2 hrs. DCs were washed and cultured with 1×10^5 naïve CD4 $^+$ T cells for 5 days. Cell cultures were pulsed with thymidine for the last 16 hours and incorporation of radionucleotide was measured by β -scintillation
10 spectroscopy.

Fig. 6 depicts GM-CSF+IL-15 DCs inducing a distinct cytokine pattern in naïve, allogeneic CD4 $^+$ T cells. Non-activated or activated GM-CSF+IL-15-derived DCs (5×10^3) were co-cultured with CD4 $^+$ T cells (1×10^5) for 5 days. The supernatants were collected, and the cells were stimulated with phytohaemagglutinin (PHA) for 24 hours.
15 The supernatants were collected and assayed for cytokines using ELISA kits. Each point represents one independent experiment. The histograms represent the means of all experiments.

Fig. 7A and 7B depict the fact that GM-CSF+IL-15-derived DCs induce recall immune responses to viral antigen. HLA-A201+DCs were activated with a mix of
20 macrophage cytokines (GM-CSF/IL-1/TNF- α /IL-6) or CD40L and pulsed for 16 hours with influenza-matrix peptide (Flu-MP). They were then harvested and used as stimulatory cells of autologous CD8 $^+$ T cells. Flu-MP antigen-specific effector CD8 $^+$ T cells were expanded for 14 days with restimulation with peptide-loaded DCs after the first week of culture. Effector cells were harvested and assayed for their ability to kill T2 cells
25 (TAP-deficient HLA-A201+ hybridoma) unpulsed (negative control) or pulsed with Flu-MP peptide in a standard chromium release assay. Specific lysis was determined at 1:1, 1:2.5, 1:5, 1:1.75, and 1:30 E:T ratio (Fig. 7A). Effector cells were collected and assayed for number of IFN- γ -secreting cells per 100,000 effectors. Data represent the number of IFN γ $^+$ cells responding to Flu-MP peptide-pulsed T2 cells (Fig. 7B).

Fig. 8 depicts the lack of proliferation of cells obtained by culturing monocytes purified by the negative depletion protocol in the presence of IL-15 alone and IL-4 alone, and the proliferation of cells obtained by culturing cells obtained by the adherent protocol in the presence of IL-15 alone and IL-4 alone. Cells obtained by both protocols were
5 grown in the presence of IL-15 alone or IL-4 alone for five days. Afterwards, the cells were cultured in a 96 well plate for an additional 10 hours in the presence of ^3H -thymidine. Thymidine uptake was then measured via beta counter.

Fig. 9A, 9B, 9C, 9D, 9E and 9F depict time kinetic studies which indicate that the cells obtained by culturing purified monocytes in the presence of IL-15 are not immature
10 dendritic cells. Monocytes purified by negative depletion were cultured in 6 well plates in the presence of IL-15. On Day 2 (Fig. 9A and 9B), Day 4 (Fig. 9C and 9D), and Day 6 (Fig. 9E and 9F), an aliquot of the cells was harvested and stained with anti-HLA-DR, anti-CD1a, anti-CD14, anti-CD40, anti-CD80, anti-CD83, and anti-CD86 and analyzed by flow cytometry.

Fig 10A and 10B depict IL-15 in combination with GM-CSF can induce
15 monocytes from blood of patients with stage IV melanoma to differentiate into dendritic cells under conditions approved for the clinical use of generated cells, i.e, in X-VIVO medium supplemented with 10% autologous serum. The induced dendritic cells can be further activated/induced to mature by a mix of macrophage cytokines (GM-CSF/IL-
20 1/TNF- α /IL-6) all of which are approved for generation of cells to be injected to patients.

DETAILED DESCRIPTION OF THE INVENTION

The present invention discloses compositions and methods of producing immature dendritic cells, activated dendritic cells, and mature dendritic cells, more preferably antigen-presenting dendritic cells, even more preferably of human origin, using
25 interleukin-15 (IL-15). This invention resides, generally, in the use of IL-15 in combination with a growth factor, preferably GM-CSF, to cause or stimulate the differentiation or production of fully functional dendritic cells, immature dendritic cells, and/or activated dendritic cells from precursors thereof in vitro, ex vivo or in vivo. This

invention also provides for the use of IL-15 alone to cause or stimulate the differentiation or production of precursor dendritic cells from monocytes in vitro, ex vivo or in vivo.

As demonstrated herein, the combination of IL-15 and GM-CSF is able to cause, induce, or stimulate the differentiation of monocytes and circulating DC precursors into dendritic cells ("DCs"). Enriched adherent blood CD14⁺ cells cultured for six days with GM-CSF and IL-15 differentiated into typical immature CD1a⁺HLA-DR⁺CD14⁺ DCs. As opposed to cultures made with GM-CSF+IL-4, 5-10% of GM-CSF+IL-15-derived DCs (IL-15 DCs) express Langerhans cell-specific markers (e.g., LAG-Langerin and CCR6). Langerhans cells (LCs) from the skin and other mucosal epithelia represent a subset of immature DCs that migrate to the draining lymph node after capture of antigens from invading pathogens. Such LCs subset may be removed from the composition by depletion, if desired.

The results presented herein further disclose that immature DCs obtained by using IL-15 in combination with GM-CSF efficiently capture soluble and particulate antigens as demonstrated using FITC-Dextran and dying-cell bodies. Furthermore, agents such as LPS, TNF- α and CD40L diminish the IL-15 DCs' antigen-capturing capacity and promote their maturation (CD83⁺ DC-LAMP⁺). These DCs are also able to induce the proliferation of allogeneic CD4⁺ and CD8⁺ T cells and are able to process or present soluble (TT) and particulate (dying cell bodies) antigens to autologous T lymphocytes. Moreover, the mature DCs, loaded with Flu-MP peptide, are able to induce the expression of INF γ -producing CD8⁺ T cells and peptide-specific CTLs. CD4⁺ T cells cultured in the presence of these DCs produce high levels of IFN γ , low IL-10, and no IL-4, indicating Th₁-type skewing. Taken together, these results thus demonstrate a novel role for IL-15 in the generation of DCs, particularly LCs.

Interleukin-15 (IL-15) is a known cytokine, whose isolation, cloning and sequence were reported by Grabstein et al (U.S. Patent No. 5,747,024; and Grabstein et al. 1994. *Science* 264:965-968) and is presented in SEQ ID NO:1 (amino acid sequence given in SEQ ID NO:2). IL-15 uses the IL-2R γ chain common to IL-2, IL-4, IL-7, IL-9 and IL-13, and various properties of IL-15 have been described in the art, such as a potent adjuvant

effect when administered in combination with a vaccine antigen as disclosed in U.S. Patent No. 5,747,024. Within the context of the present invention, IL-15 refers to the polypeptide or corresponding nucleic acid as disclosed in SEQ ID NO:1, as well as any variant thereof, including any naturally-occurring derivatives thereof or homologs isolated
5 from various mammalian species. Variants more generally encompass any polypeptide having one or several amino acid modifications as compared to SEQ ID NO:1, including mutation(s), deletion(s), insertion(s), substitution(s), alone or in various combinations(s). More particularly, variants as used herein are any polypeptide having the property to stimulate or induce production of dendritic cells from monocytes as disclosed herein,
10 encoded by a nucleic acid sequence hybridizing, under conventional, moderate stringency, with the nucleic acid sequence disclosed in SEQ ID NO:1. Still preferably, IL-15 has at least 80% identity with the amino acid sequence disclosed in SEQ ID NO:2.

IL-15 is more preferably a recombinant IL-15, i.e., produced by expression, in any suitable host cell, of a nucleic acid molecule encoding such a polypeptide. Suitable host
15 cells include, for example, mammalian cells, prokaryotic cells, or yeast cells. In this regard, recombinant human IL-15 is commercially available from various sources, including R&D Systems, Inc.(Minneapolis, MN, US). A preferred IL-15 for use in the present invention is a human IL-15, even more preferably a recombinant human IL-15. For use in the present invention, although essentially pure IL-15 preparations are
20 preferred, it should be understood that any IL-15 preparation may be used, in combination with other factors, diluents, adjuvants, biological fluids, etc. Furthermore, while the process preferably utilizes an IL-15 polypeptide, the invention also includes the use of any IL-15-encoding nucleic acid molecule. In particular, the precursor cells may be contacted directly with a nucleic acid coding IL-15 incorporated in a suitable expression vector, or
25 with a cell population containing an IL-15-encoding nucleic acid molecule and expressing IL-15. Such cells may further produce a growth factor such as GM-CSF. For in vivo production, the patient or subject may be administered with either IL-15 or any nucleic acid encoding IL-15 in a suitable expression vector. The concentration of IL-15 may be adjusted by the skilled artisan.

For the performance of the present invention, DC precursors are preferably contacted or treated with IL-15 and a growth factor, preferably GM-CSF, in vitro, ex vivo or in vivo. DC precursors include any cell or cell population which is capable of differentiating into dendritic cells in vitro, ex vivo or in vivo, upon treatment according to the present invention. Typical DC precursors include monocytes and circulating DC precursors. For in vitro or ex vivo uses, DC precursors may be obtained from various biological materials, including blood and bone marrow, preferably blood. Typically, DC precursors comprise peripheral blood mononuclear cells (PBMC) or peripheral blood monocytes isolated therefrom. PBMC can be prepared by conventional aphaeresis, and monocytes isolated by Ficoll gradient, optionally coupled to depletion of T cells, B cells, and/or NK cells. Exemplary methods of isolating DC precursors useful in the present invention have been reported by Inaba et al (Inaba et al. 1992. *J Exp Med* 175:1157) and Romani et al (Romani et al. 1994. *J Exp Med* 180:83).

For in vitro and ex vivo applications, DC precursors may be contacted with IL-15, and optionally a growth factor such as GM-CSF, in any appropriate device, such as plate, well, flask, tube, etc., and preferably under sterile conditions. Typically, between 1×10^4 to 1×10^8 cells, more preferably between 1×10^5 to 1×10^7 cells are used, depending on the amount of DC precursors available or collected. The amount of cytokine and/or growth factor such as GM-CSF may be adjusted by the skilled person. Typically, IL-15 is used at a concentration of about 50 ng/ml to about 500 ng/ml, and more preferably about 100 ng/ml to about 300 ng/ml. GM-CSF may be employed at a concentration of about 10 ng/ml to about 300 ng/ml, and more preferably about 50 ng/ml to about 200 ng/ml. Similar concentrations or doses may be used for in vivo induction or stimulation of dendritic cell production or differentiation. The contacting cells may be contacted in vitro with the above reagents for about 24 hours to 15 days. Usually, immature DCs are produced and collected at Day 3-Day 7 after stimulation of DC precursors, preferably between 4-6 days post-treatment. Typically, 1×10^6 DC precursor cells (e.g., monocytes) yield about 1×10^3 immature DCs. Immature DCs obtained are particularly characterized by the expression of CD1a and HLA-DR, the absence of CD14, and minimal levels or no

expression of DC-LAMP (DC-lysosomal-associated membrane glycoprotein), CD40, CD80 and CD83.

Immature DCs may be collected and maintained in any suitable culture media, such as RPMI (GIBCO BRL, Grand Island, NY, US), X-VIVO 15 (BioWhittaker, Inc.,
5 Walkersville, MD, US), AIMV (GIBCO BRL, Grand Island, NY, US), and DMEM (GIBCO BRL, Grand Island, NY, US), etc. The cells may be stored or frozen under conventional conditions. Quality, purity and lack of contamination may be assessed using conventional methods known in the art, such as antibody-labeling, dye labeling, etc.

A preferred aspect of this invention resides in a method for inducing or stimulating
10 dendritic cell differentiation from precursors thereof, more particularly monocytes, comprising contacting the precursors with a combination of IL-15 and a growth factor, preferably GM-CSF, in vitro, ex vivo or in vivo. In a specific embodiment of this invention, immature dendritic cells or activated dendritic cells are produced from precursors of dendritic cells such as monocytes by contacting the precursors with a
15 combination of IL-15 and a growth factor, preferably GM-CSF, in vitro, ex vivo or in vivo.

Immature DCs have utility in many applications, including diagnostic, therapy, vaccination, research, screening, gene delivery, etc. In particular, because of their potent immunogenic properties, DCs may be sensitized to an antigen and used to modulate an
20 immune reaction or response in vivo, for instance to induce antigen-specific tolerance. As used herein, the modulation of the immune response by DCs includes increases, decreases, or changes to other types of immune responses.

Activated DCs have utility in many applications, including diagnostic, therapy, vaccination, research, screening, gene delivery, etc. In particular, because of their potent
25 immunogenic properties, DCs may be sensitized to an antigen and used to cause, induce, or stimulate an immune reaction or response in vivo. They may also be used to produce, screen, or expand, in vitro or ex vivo, antigen-specific cytotoxic T lymphocytes (CTL).

In this respect, DCs may be sensitized to one or several antigens according to various techniques known in the art, such as by placing the DCs in contact with an antigen

(“antigen pulsing”), an antigenic peptide (“peptide pulsing”), an antigenic protein complex, cells or cell membranes expressing antigens or antigenic peptides, texosomes, liposomes containing antigens or antigenic peptides, nucleic acids encoding antigens or antigenic peptides (possibly incorporated in plasmids or viral vectors), or total RNAs from
5 a tumor cell. These methods have been disclosed, for instance, in International Application No. WO99/03499. Antigens may be tumor antigens, viral antigens, bacterial antigens, etc., more generally, any peptide or polypeptide against which an immune response or reaction is sought. The term “sensitized” indicates that the antigen or a portion thereof is exposed at the surface of the DCs, preferably in complex with molecules
10 of the major histocompatibility complex (MHC).

Immature dendritic cells may also be used to produce membrane vesicles (also termed dexosomes), as described in WO99/03499, prior to or after antigen sensitization. In this regard, a particular object of this invention concerns a method of producing membrane vesicles, comprising (1) providing immature dendritic cells by contacting
15 precursors thereof, more particularly monocytes, with IL-15 and GM-CSF, (2) culturing the dendritic cells under conditions allowing the release of membrane vesicles, and (3) isolating the membrane vesicles. Even more preferably, the method comprises, prior to Step (2), a sensitization of the DCs to one or several antigens. In Step (1), the DC precursors are preferably contacted with a combination of IL-15 and GM-CSF, as
20 described herein. In Step (2), the DCs may be cultured in any conventional media, preferably in the presence of a stimulating compound and/or treatment and/or environment, to increase membrane vesicle release. Preferred stimulating conditions include culturing in the presence of cytokine(s), more preferably IL-10, gamma interferon (IFN- γ), or IL-12, irradiation and/or at low pH (e.g., pH=5), as described in WO99/03499.
25 The membrane vesicles (or dexosomes) may be recovered, for instance, by centrifugation and/or chromatographic methods.

As indicated above, these membrane vesicles or dendritic cells can be used to modulate an immune response in a patient, in particular a mammal, more preferably a human. In this regard, the invention also relates to a method of modulating an immune
30 response in a patient, comprising administering to the patient an effective amount of

dendritic cells prepared in the presence of IL-15 or membrane vesicles derived therefrom. Even more preferably, the dendritic cells and/or membrane vesicles are sensitized to one or several antigens. In a specific embodiment, the DCs have been prepared from blood precursors, cultured in the presence of IL-15 and preferably a growth factor such as GM-CSF.

Another aspect of this invention lies in a method of modulating an immune response in a patient, via inducing dendritic cells in a patient, comprising the concurrent or sequential administration to the patient of an effective amount of IL-15 and a growth factor such as GM-CSF and/or maturation factor (e.g., CpG nucleotides or type I interferon). Sequential administration indicates that the compounds may be injected separately, in any order. Preferred administration routes include intravenous, intra-arterial, intra-muscular, intra-dermal, and local (e.g., intra-tumoral or at the vicinity of a tumor site).

The DCs prepared according to the above methods may furthermore be activated or cultured in vitro or ex vivo under conditions allowing their maturation. In this respect, an object of the present invention resides in a method of producing mature DCs, comprising (1) preparing immature dendritic cells by contacting precursors thereof, more particularly monocytes, with IL-15 in the presence of a growth factor such as GM-CSF and (2) culturing the dendritic cells under conditions allowing their maturation, preferably in the presence of lipopolysaccharide (LPS), CD40L or poly(I):(C). Mature DCs show elevated expression of CD80, CD83 and CD86. Mature DCs may be sensitized to antigens, either prior, during, or after maturation.

Another object of the present invention resides in a method of producing antigen-specific CTL in vitro or ex vivo, comprising contacting antigen-sensitized dendritic cells as described above with a population of cells comprising T lymphocytes and recovering the activated CTLs which are specific for the antigen. The population comprising T lymphocytes may be isolated CD8⁺ T lymphocytes or peripheral blood mononuclear cells. Activated CTLs may be recovered and selected by measuring clonal proliferation, target cell lysis and/or cytokine release, as described in the examples provided herein. The antigen-specific CTLs may be further expanded by culture in the presence of antigen-

sensitized DCs. This method is also applicable to the production of activated CD4+ T lymphocytes. Furthermore, the method may also be performed in vivo, to activate or increase a CTL response or a CD4+ response to an antigen in vivo.

5 The invention also includes compositions of matter comprising dendritic cells or membrane vesicles derived therefrom, obtainable as disclosed above. In particular, the invention relates to compositions comprising dendritic cells obtained by contacting in vitro or ex vivo precursors thereof in the presence of IL-15 and a growth factor such as GM-CSF, more particularly immature dendritic cells, even more particularly human immature dendritic cells. These dendritic cells may be sensitized to one or several
10 antigens, as described above.

The invention also relates to compositions comprising membrane vesicles obtained from immature dendritic cells obtained by contacting in vitro or ex vivo precursors thereof in the presence of IL-15 and a growth factor such as GM-CSF. The invention further relates to compositions comprising IL-15 and GM-CSF, for concurrent or sequential use.
15 The compositions may comprise any pharmaceutically acceptable carrier or excipient, e.g., buffer, saline solution and adjuvants. They can be used to induce or stimulate an immune response or reaction in vivo, e.g., for treating a patient with tumor or immune disease such as allergy, asthma, and/or autoimmune diseases. They are particularly suited for treating cancer in a patient.

20 Another specific embodiment of this invention resides in a method of producing precursor dendritic cells from monocytes, comprising contacting the monocytes with IL-15 in vitro, ex vivo or in vivo. The invention also includes compositions of matter comprising precursor dendritic cells derived by contacting monocytes with IL-15 in vitro, ex vivo or in vivo.

25 Monoclonal antibodies and recombinant growth factors and reagents

Monoclonal antibodies (mAbs) used in this study were: CD14, HLA-DR (Becton Dickinson, Franklin Lakes, NJ, US.); CD86, (PharMingen, San Diego, CA, US); CD40, HLA-AB, CCR6 (R&D Systems, Inc., Minneapolis, MN, US), CD1a (Dako Corp., Carpinteria, CA, US), CD80, and CD83 (Coulter/Immunotech, Fullerton, CA, US).

Recombinant human cytokines used in this study were: rhGM-CSF (100 ng/ml, Leukine, Immunex Corp., Seattle, WA, US); rhIL-4 (20 ng/ml, R&D Systems, Inc., Minneapolis, MN, US); rhIL-15 (200 ng/ml, R&D Systems, Inc., Minneapolis, MN, US); TGF- β 1 (10 ng/ml, R&D Systems, Inc., Minneapolis, MN, US); and huCD40L (Immunex Corp., Seattle, WA, US).

Complete RPMI medium consisted of RPMI 1640, 1% L-Glutamine, 1% penicillin/streptomycin, 50mM 2-ME, 1% sodium-pyruvate, 1% essential amino acids and heat inactivated 10% fetal calf serum (all from GIBCO BRL, Grand Island, NY, US). Poly(I):(C) was purchased from Pharmacia (Piscataway, NJ, U.S.A.), and lipopolysaccharide (LPS) was purchased from Sigma Chemical Co. (St. Louis, MO, US).

Cell culture and phenotypic analysis of monocytes derived dendritic cells

Peripheral blood monocytes were isolated by immunomagnetic depletion (Dynabeads; Dynal Biotech, Lake Success, NY, US) after separation of blood mononuclear cells followed by a Ficoll-PaqueTM gradient. The pellet was recovered for T cell purification, and monocytes were isolated negatively by positive magnetic depletion of T-cells, B-cells, and natural killer (NK) cells using purified anti-CD3, anti-CD16, anti-CD19, anti-CD56, and anti-glycophorin A antibodies. Afterwards, enriched CD14⁺ monocytes were cultured in 6 well plates (1 x 10⁶ cells/well). These cells were cultured for 6 days in the presence of 100 ng/ml GM-CSF and 200 ng/ml IL-15 (GM-CSF+IL-15), or 100 ng/ml GM-CSF and 20 ng/ml IL-4 (GM-CSF+IL-4). On Day 6, cells were harvested and stained with anti-CD14-PE, anti-CD83-PE, anti-HLA-DR-PerCP, and anti-CD1a-FITC. In some experiments, DCs were activated with LPS (300 ng/ml), and their phenotypes were analyzed. The analysis was performed by FACS Calibur.

Endocytotic activity of CD1a⁺ DC

The endocytotic activity of DCs was determined as follows. Activated with LPS (300 ng/ml) or non-activated GM-CSF+IL-15-derived DCs were harvested on Day 6 and incubated with 100 μ g/ml FITC-Dextran for 30 min at 37°C. As a control, part of the DCs was incubated with the same amount of FITC-Dextran on ice. The cells were washed with cold phosphate buffered saline (PBS) with 10% fetal calf serum and analyzed by FACS Calibur or confocal microscopy.

Confocal Microscopy

Intracellular immunofluorescence staining was conducted as previously described (de Saint Vis, et al. 1998. *Immunity* 9:325-336; Valladeau et al. 2000. *Immunity* 12:71-81). On polylysine-coated coverslips, cells were fixed for 15 min with 4% paraformaldehyde in PBS. Afterwards, these cells were washed twice in 10 mM glycine in PBS and twice in PBS, and permeabilized with PBS containing 0.5% saponin and 1% bovine serum albumin (BSA) for 30 min. Coverslips were incubated for 30 min at room temperature with 5 μ g/ml anti-DC-LAMP or anti-Langerin (gifts from Drs. Lebecque and Saeland, Schering-Plough, Dardilly, France). After three washes, cells were incubated for 30 min with secondary labeled donkey anti-mouse antibodies coupled to Texas red (Jackson Immuno Research Laboratories, Inc., West Grove, PA, US), washed, incubated with mouse pre-immune serum for 30 min, washed again, incubated for 30 min with a second primary antibody anti-HLA-DR directly coupled to fluorescein (Becton Dickinson, Franklin Lakes, NJ, US). Coverslips were mounted onto glass slides with Fluoromount (Southern Biotechnology Associates, Birmingham, AL, US). Confocal microscopy was performed using a TCS SP microscope equipped with argon and krypton ion lasers (Leica Microsystem, Heidelberg, Germany).

T-cell proliferation

DCs cultured in GM-CSF+IL-15, GM-CSF+IL-4, or GM-CSF+IL-4/TGF- β 1 were harvested, washed with complete RPMI and cultured with 1×10^5 freshly isolated CD4⁺ or CD8⁺ allogeneic T cells derived from peripheral blood. In some experiments, DCs were activated with poly(I):(C) (6.5 μ g/ml) for 16 hours. DC-T cell cultures were set up for 5 days in complete RPMI plus 10% human AB serum. Cells were pulsed for the last 16 hours with 0.5 mCi [³H]thymidine per well, and incorporation of the radionucleotide was measured by β -scintillation spectroscopy. To assay autologous T cell proliferation, GM-CSF+IL-15 or GM-CSF+IL-4 DCs were collected, washed, and then pulsed with tetanus toxoid (TT) for 48 hours. Afterwards, DCs were washed three times with PBS and then resuspended in complete RPMI plus 10% human AB serum. Cells were added in triplicate at various concentrations to 1×10^5 autologous T cells/well in 96-well tissue culture plates (Falcon, Oxnard, CA, US). To assay capture of dying cell bodies by DCs, DCs were

incubated for two hours with Me275 cell line bodies, which were prepared by treating with betulinic acid (10 µg/ml, Aldrich, Milwaukee, WI, US) for 48 hours. Afterwards, DCs were sorted based on forward/side scatter using a FACS Vantage. DCs were then co-cultured with autologous CD4⁺ T cells. CD4⁺ or CD8⁺ T cells were isolated by the
5 standard Ficoll-paque procedure followed by magnetic depletion of non-T cells (Dynabeads; Dynal Biotech, Lake Success, NY, US).

Cytokine analysis

For cytokine analysis, supernatants were harvested 5 days after co-culture of GM-CSF+IL-15-derived DCs/CD4⁺ T cells, and the cells were re-stimulated with PHA (10
10 µg/ml) in fresh medium for 24 hours. Afterwards, the supernatants were collected and assayed for cytokine release by utilizing ELISA kits from R&D Systems (Minneapolis, MN, US).

Generation of antigen-specific CTL

GM-CSF+IL-15-derived or GM-CSF+IL-4-derived DCs were stimulated with
15 CD40L or with a mix of macrophage cytokines (GM-CSF/TNF-α/IL-1/IL-6). DCs were then washed and pulsed with 10 µg/ml of Flu-MP⁵⁸⁻⁶⁶ for further 18 hours. CD8⁺ effector T cells (1 x 10⁶) were isolated by magnetic beads separation and co-cultured with either non-pulsed DCs or with Flu-MP-pulsed DCs in 1 ml complete RPMI plus 10% human AB serum. All CTL cultures received IL-7 (10 ng/ml) at the establishment of culture and IL-2
20 (10 UI/ml) on Day 7 of culture. CTL were grown for 14 days prior to assay. CTL were used for cytotoxicity assay on Day 14. Effector cells (30 x 10³/well) were plated in 96-well round-bottom plates with T2 (1 x 10³/well) target cells incubated overnight with Flu-MP⁵⁸⁻⁶⁶ and labeled with ⁵¹Cr (Amersham Pharmacia, Biotech, Inc., Piscataway, NJ, US). After 4 hours, supernatants were harvested using a harvesting frame and released
25 chromium-labeled protein was measured using γ-counter (Packard Instruments Co, Meriden, CT, US). Percentage of antigen-specific lysis was then determined.

INF-γ Enzyme-linked Immunospot assay

To determine antigen-specific, IFN-γ-producing effector T cells, an enzyme-linked immunospot (ELISPOT) assay was used as described by Dhodapkar, et al. (Dhodapkar, et

al. 1999. *J Clin Invest* 104:173-180). CD8⁺ T cells (1×10^5 /well) were added in triplicate to nitrocellulose-bottomed 96-well plates (MAHA S4510; Millipore Corp., Bedford, MA, US) precoated with the primary anti-IFN- γ mAb (1-D1K; Mabtech AB, Nacka, Sweden) in 50 μ l complete RPMI/well. For the detection of specific reactive T cells, autologous
5 mature monocyte-derived DCs pulsed with MHC class I-restricted-peptides (Flu-MP) were added at 1×10^4 /well (final volume 100 μ l/well). After incubation for 20 hours, wells were washed 6 times, incubated with biotinylated secondary mAb against IFN- γ (7B6-1; Mabtech AB) for 2 hours, washed and stained with Vectastain Elite kit (Vector Laboratories, Inc., Burlingame, CA, US). Responses were counted as positive if a
10 minimum of 10 spot forming cells (SFC)/ 2×10^5 cells were detected and if the number of spots were at least twice that of the negative control wells.

Comparison of the monocyte source in response to the presence of IL-15 alone

To determine if the monocyte source can affect the results obtained when the monocytes are treated with IL-15 alone, monocytes were obtained by the following two
15 protocols: (1) negative depletion by magnetic beads and (2) an adherent protocol. In the negative depletion protocol, peripheral blood monocytes were isolated by immunomagnetic depletion (Dynabeads, Dynal Biotech, Lake Success, NY, US) after separation of blood mononuclear cells followed by a Ficoll-PaqueTM gradient. The pellet was recovered for T cell purification and monocytes were isolated negatively by positive
20 magnetic depletion of T cells, B cells, and NK cells using purified anti-CD3, anti-CD16, anti-CD19, anti-CD56 and anti-glycophorin A antibodies. Afterwards, enriched CD14⁺ monocytes were cultured in 6 well plates (1×10^6 /well). An aliquot of these cells was cultured for 6 days in the presence of IL-15 alone and IL-4 alone, respectively. On Day 2, Day 4, and Day 6, cells were harvested, stained with anti-CD14, anti-CD83, anti-CD80,
25 anti HLA-DR, anti-CD1a, anti-CD40, and anti-CD86, and analyzed by FACS Calibur. In the adherent protocol, crude lymphocytes were cultured in 6-well plates for 2 hours, and the cells were washed three times with PBS containing 2% FCS to remove non-adherent cells. The adherent cells were then cultured with IL-15 alone and with IL-4 alone. Labeled thymidine was added to cells obtained by both the negative depletion protocol
30 and the adherent protocol, and thymidine uptake was measured on Day 6 via β -counter.

Additional aspects and advantages of the present invention will be described in the following examples, which should be regarded as illustrative and not limiting the scope of the present application.

Example 1: Monocytes cultured with GM-CSF+IL-15 yield immature DCs (IL-15 DCs)

5 A study was conducted to determine whether IL-15 may be involved in the differentiation of immature DC and most particularly LCs. Monocytes were highly purified by negative depletion using magnetic beads (98%) and cultured with IL-15 in combination with GM-CSF. After six days of culture with medium containing GM-CSF+IL-15, characteristic phase contrast morphology of IL-15 DCs indicated extensive
10 cell aggregation. Similar cell aggregation was observed in cultures containing a combination of GM-CSF+IL-4. Representative phase contrast and Giemsa staining of the IL-15 DCs revealed extending veils, a morphology suggestive of DCs. Flow cytometry analysis showed that all the cells had acquired CD1a, lost CD14 and expressed relatively high levels of HLA-DR (Fig. 1A). Thus, monocytes cultured with GM-CSF+IL-15
15 differentiated into cells with a phenotype of immature DCs. Monocytes required 4-6 days of culture to fully differentiate into immature DCs (Fig. 1B) (hereinafter these cells are referred to as IL-15-DCs). After six days, 1×10^6 monocytes yielded a mean 0.53×10^6 immature DCs (mean of 5 experiments, SD=176.86). The generation of IL-15-derived DCs was independent of endogenous IL-4 production as determined by daily additions of
20 neutralizing anti-IL4. Such treatment prevented the differentiation of monocytes into DCs in response to GM-CSF+IL-4, but not in response to GM-CSF+IL-15, while addition of anti-IL-15 or anti-IL-2R γ chain blocked significantly the generation of CD1a⁺ DCs in response to GM-CSF+IL-15.

Example 2: IL-15 DCs can be activated into mature DCs

25 As shown in Fig. 2A, IL-15-derived DCs express HLA-ABC Class I, low levels of CD86 and minimal levels CD40, CD80, CD83. Upon activation with LPS, cells show increased expression of CD40, CD80, CD86, and CD83. Confocal microscopy sections revealed the accumulation of intracellular DC-LAMP in the GM-CSF/IL-15 condition, a marker of mature DCs. The maturation of IL-15-derived DCs could be induced equally
30 well with LPS and CD40L. At variance with DCs generated with IL-4, IL-15-derived

DCs always display some activated cells as shown by spontaneous expression of DC-LAMP in 5-10% of the cells. As immature DCs, IL-15-derived DCs were capable of efficiently capturing antigens, e.g., FITC-Dextran (Fig. 2B). However, they lost this capacity upon maturation e.g., with LPS (Fig. 2A).

5 Example 3: IL-15 DCs include LCs

CD34⁺ hematopoietic progenitor cells cultured with GM-CSF+TNF- α yield both interstitial DCs and LCs. Monocytes cultured with GM-CSF+IL-4 did not differentiate into LCs unless TGF- β 1 was added to these cultures. To date, two markers distinguish LCs: CCR6, the receptor to MIP-3 α/β defensin, and LAG/Langerin, a lectin involved in the formation of Birbeck granules. As shown in Fig. 3A, up to 10% of IL-15-derived DCs expressed CCR6, while DCs generated with GM-CSF+IL-4 did not. Dual-immunofluorescence confocal microscopy of HLA-DR/Langerin was performed on monocyte-derived DCs cultured in vitro for 6 days with GM-CSF+IL-4, GM-CSF+IL-15, and GM-CSF+IL-4/TGF- β 1. Confocal sections revealed the accumulation of intracellular Langerin in DCs generated by culturing monocytes with GM-CSF+IL-15 and GM-CSF+IL-4/TGF- β 1 but not with GM-CSF+IL-4.

Example 4: IL-15 DCs present soluble and particulate antigens to T cells

IL-15 DCs are efficient antigen-presenting cells which can induce the proliferation of allogeneic CD4⁺ and CD8⁺ T cells (Fig. 4A and 4B). DCs generated by culturing of monocytes with either GM-CSF+IL-15, GM-CSF+IL-4, or GM-CSF+IL-4/TGF-β1 were comparable in their ability to induce allogeneic mixed lymphocyte reaction (MLR) (Fig. 4C). They were considerably more efficient than cells generated with GM-CSF alone (Fig. 4C). IL-15 DCs activated for 24 hours with poly (I):(C) (Fig. 4A, B), CD40L or LPS showed increased allo-stimulatory capacity. The capacity to present antigens to autologous CD4⁺ T cells was also assayed. Thus, IL-15 DCs loaded with soluble tetanus toxoid (TT) induced autologous CD4⁺ T cells to proliferate (Fig. 5A). Furthermore, IL-15 DCs that had captured dying cell bodies also induced the proliferation of autologous T lymphocytes (Fig. 5B).

Example 5: IL-15 DCs induce CD4⁺ T cells to produce INF- γ

The capacity of IL-15 DCs to skew T cell differentiation was assessed by co-culturing them with naïve CD4⁺CD45RA⁺ T cells for five days. Afterwards, cells were washed and stimulated with PHA, and supernatants were assayed for cytokine production.

- 5 Non-activated IL-15 DCs induced T cells to produce high amounts of IFN- γ , but not IL-4 or IL-10 (Fig. 6). Co-cultures of T cells and stimulated IL-15 DCs with Poly (I):(C) contained higher levels of IFN- γ (Fig. 6). Then, IL-10 could be observed in supernatants (Fig. 6).

Example 6: Cytotoxicity assay

- 10 The ability of IL-15 DCs to stimulate MHC class-I restricted CTL responses was evaluated. IL-15 DCs were pulsed with Flu-MP peptide and used to stimulate purified CD8⁺ T cells. After two seven-day culture cycles performed in the presence of IL-7 (both cycles) and IL-2 (second cycle), the elicited T cells were able to efficiently kill T2 cells loaded with Flu-MP peptide (55% specific lysis at a 2.5:1 ratio and 60% specific lysis at a
- 15 5:1 E:T ratio). T cell lines, which were generated by co-culture with unloaded IL-15 DCs (control DCs) induced less than 5% specific lysis (Fig. 7A). However, as shown in Fig. 7B, IL-15 DCs loaded with Flu-MP peptide were comparable to GM-CSF+IL-4 DCs in stimulating the generation of peptide-specific CTL precursors able to produce IFN- γ . Unpulsed DCs did not induce specific IFN- γ producing cells (Fig. 7B).

- 20 Example 7: Culture of monocytes in the presence of IL-15 alone

- When purified monocytes obtained by the negative depletion protocol were grown in the presence of IL-15 and GM-CSF, the resulting GM-CSF+IL-15 DCs were CD1a⁺ and CD14⁻, indicative of immature dendritic cells. However, when these monocytes were cultured with IL-15 alone, the resulting cells were CD1a⁺ and CD14⁺, indicative of
- 25 precursor dendritic cells and not immature dendritic cells. As shown in Fig. 8, the purified monocytes obtained by the negative depletion protocol did not proliferate when cultured in the presence of IL-15 or IL-4. Time kinetic studies presented in Fig. 9A-9F further demonstrate that the cells obtained after culturing these purified monocytes in IL-15 alone were not immature dendritic cells.

As shown in thymidine uptake studies illustrated in Fig. 8, when adherent cells obtained by the adherent protocol were cultured in the presence of IL-15 alone, the resulting cell culture showed evidence of proliferation. These results indicated that the adherent cells not only included monocytes but also contaminating T cells, B cells, and
5 NK cells, known to respond to IL-15 by proliferation.

As indicated above, IL15 in conjunction with GM-CSF induces highly purified monocytes to differentiate into immature DCs. The cells generated under these conditions are in many ways comparable to DCs generated with GM-CSF+IL4: 1) the same phenotype; 2) the same capacity to capture FITC-Dextran and dying cell bodies; 3) the
10 same capacity to mature in response to various stimuli; 4) the same ability to process and present soluble and particulate antigens; 5) the same ability to induce the differentiation of T cells into the Th1 pathway; and 6) the same ability to induce the differentiation of CD8 T cells into specific CTL. Two differences were observed between DCs generated in the presence of IL-4 and DCs generated in the presence of IL-15: 1) IL-15 cultures included
15 some mature DCs as shown by expression of DC-LAMP; and 2) IL-15 cultures contained Langerhans cells. This latter observation is relevant to IL-15, which is a product of KCs, the cells that fully surround the LCs within the epidermis. Thus, monocytes that extravasate from dermal vessels would differentiate into interstitial DCs when encountering the GM-CSF and IL-4 produced by dermal mast cells; but would
20 differentiate into Langerhans cells when encountering the GM-CSF and IL-15 produced by KCs.

Example 8: IL-15 induces monocytes from patients with
metastatic cancer to differentiate into dendritic cells

As shown in Fig. 10, the adherent fraction of peripheral blood mononuclear cells
25 give rise, upon culturing with IL-15 and GM-CSF to cells with the phenotype of dendritic cells including a Langerhans cell component as determined by surface expression of Langerin. These cells are obtained in culture performed under conditions approved for clinical use and the reinjection of cultured cells into patient. The cells are generated in X-VIVO culture medium supplemented with 2%-10% autologous serum.

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WE CLAIM:

1. A method for producing immature dendritic cells from dendritic cell precursors in vitro or ex vivo comprising contacting said precursors with a composition comprising interleukin-15 and at least one growth factor.
2. The method of Claim 2, further comprising sensitizing said immature dendritic cells to an antigen.
3. A method for modulating an immune response in a patient comprising administration of the immature dendritic cells of Claim 1 or 2 to said patient.
4. A method for producing activated dendritic cells from dendritic cell precursors in vitro or ex vivo comprising contacting said precursors with a composition comprising interleukin-15 and at least one growth factor.
5. The method of Claim 4, further comprising sensitizing said activated dendritic cells to an antigen.
6. A method for modulating an immune response in a patient comprising administration of said activated dendritic cells of Claim 4 or 5 to said patient.
7. A method for producing mature dendritic cells from dendritic cell precursors in vitro or ex vivo comprising contacting said precursors with a composition comprising interleukin-15 and at least one growth factor to form immature dendritic cells and culturing said immature dendritic cells in the presence of an maturation agent and under
5 conditions to promote maturation of said immature dendritic cells to form mature dendritic cells.
8. The method of Claim 7, further comprising sensitizing said dendritic cells to an antigen either prior to, during or after maturation.
9. The method of Claim 7 or 8, wherein said maturation agent is selected from the group consisting of lipopolysaccharide, CD40L, and poly(I):(C).
10. A method for modulating an immune response in a patient comprising administration of the mature dendritic cells of Claim 7, 8 or 9 to said patient.

11. A method for producing isolated membrane vesicles comprising contacting dendritic cell precursors with a composition comprising interleukin-15 and at least one growth factor to form immature dendritic cells, culturing said immature dendritic cells in the presence of at least one stimulation factor to promote the release of membrane
5 vesicles; and isolating said membrane vesicles.

12. The method of Claim 11, further comprising sensitizing said immature dendritic cells to an antigen prior to or after membrane vesicle release.

13. The method of Claim 11 or 12, wherein said stimulation factor is selected from the group consisting of a suitable cytokine, irradiation, and low pH.

14. A method for modulating an immune response in a patient comprising administration of the membrane vesicles of Claim 11, 12 or 13 to said patient.

15. A method for producing immature dendritic cells from dendritic cell precursors in a patient comprising concurrent or sequential administration of interleukin-15 and at least one growth factor to said patient.

16. A method for producing activated dendritic cells from dendritic cell precursors in a patient comprising concurrent or sequential administration of interleukin-15 and at least one growth factor to said patient.

17. A method for producing mature dendritic cells from dendritic cell precursors in a patient comprising concurrent or sequential administration of interleukin-15, at least one growth factor, and at least one maturation agent.

18. The method of Claim 17, wherein said maturation agent is selected from the group consisting of CpG oligonucleotides and type I interferon.

19. A method for modulating an immune response in a patient via dendritic cells comprising concurrent or sequential administration of interleukin-15 and at least one growth factor to said patient.

20. The method of Claim 19, further comprising concurrent or sequential administration of at least one maturation factor to said patient.

21. A method for inducing or stimulating dendritic cell differentiation from precursors thereof comprising contacting said precursors with interleukin-15 and at least one growth factor.

22. A method for producing antigen-specific cytotoxic T lymphocytes in vitro or ex vivo comprising providing antigen-sensitized dendritic cells prepared by contacting precursors thereof with interleukin-15 and at least one growth factor and subsequent sensitizing of said dendritic cells to said antigen, and contacting said antigen-sensitized
5 dendritic cells with a population of cells comprising T lymphocytes to form activated antigen-specific cytotoxic T lymphocytes.

23. A method for increasing an immune response in a patient comprising administration of said antigen-specific cytotoxic T lymphocytes of Claim 22 to said patient.

24. A method for producing antigen-specific CD4⁺ T lymphocytes in vitro or ex vivo comprising providing antigen-sensitized dendritic cells prepared by contacting precursors thereof with interleukin-15 and a growth factor and subsequent sensitizing of said dendritic cells to said antigen and contacting said antigen-sensitized dendritic cells
5 with a population of cells comprising T lymphocytes to form activated antigen-specific CD4⁺ T lymphocytes.

25. A method for modulating an immune response in a patient comprising administration of said antigen-specific CD4⁺ T lymphocytes of Claim 24 to said patient.

26. A method for producing dendritic cell precursors comprising contacting monocytes in vitro or ex vivo with interleukin-15 to form dendritic cell precursors.

27. A composition comprising immature dendritic cells produced according to the method of Claim 1 or 2.

28. A composition comprising activated dendritic cells produced according to the method of Claim 4 or 5.

29. A composition comprising mature dendritic cells produced according to the method of Claim 7 or 8.

30. A composition comprising membrane vesicles produced according to the method of Claim 11, 12 or 13.

31. A composition comprising antigen-specific cytotoxic T lymphocytes produced according to the method of Claim 22.

32. A composition comprising antigen-specific CD4+ T lymphocytes produced according to the method of Claim 24.

33. A composition comprising dendritic cell precursors produced according to the method of Claim 26.

34. The method of Claim 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, 13, 14, 15, 16, 17, 18, 19, 20, 21, 22, 23, 24, 25 or 26, wherein said interleukin-15 is in a form selected from the group consisting of isolated interleukin-15, a recombinant interleukin-15 polypeptide, a nucleic acid encoding interleukin-15, either wild type or engineered, in a suitable
5 expression vector, and a cell population expressing a nucleic acid encoding interleukin-15.

35. The composition of Claim 27, 28, 29, 30, 31, 32 or 33, wherein said interleukin-15 is in a form selected from the group consisting of isolated interleukin-15, a recombinant interleukin-15 polypeptide, a nucleic acid encoding interleukin-15, either wild type or engineered, in a suitable expression vector, and a cell population expressing a
5 nucleic acid encoding interleukin-15.

36. The method of Claim 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, 13, 14, 15, 16, 17, 18, 19, 20, 21, 22, 23, 24, 25 or 26, wherein said growth factor is in a form selected from the group consisting of isolated growth factor, a recombinant growth factor polypeptide, a nucleic acid encoding a growth factor in a suitable expression vector, and a cell population
5 expressing a nucleic acid encoding a growth factor.

37. The composition of Claim 27, 28, 29, 30, 31, 32 or 33, wherein said growth factor is in a form selected from the group consisting of isolated growth factor, a recombinant growth factor polypeptide, a nucleic acid encoding a growth factor in a suitable expression vector, and a cell population expressing a nucleic acid encoding a
5 growth factor.

38. The method of Claim 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, 13, 14, 15, 16, 17, 18, 19, 20, 21, 22, 23, 24, 25 or 26, wherein said growth factor is granulocyte-macrophage colony stimulating factor.

39. The composition of Claim 27, 28, 29, 30, 31, 32 or 33, wherein said growth factor is granulocyte-macrophage colony stimulating factor.

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Figure 1A

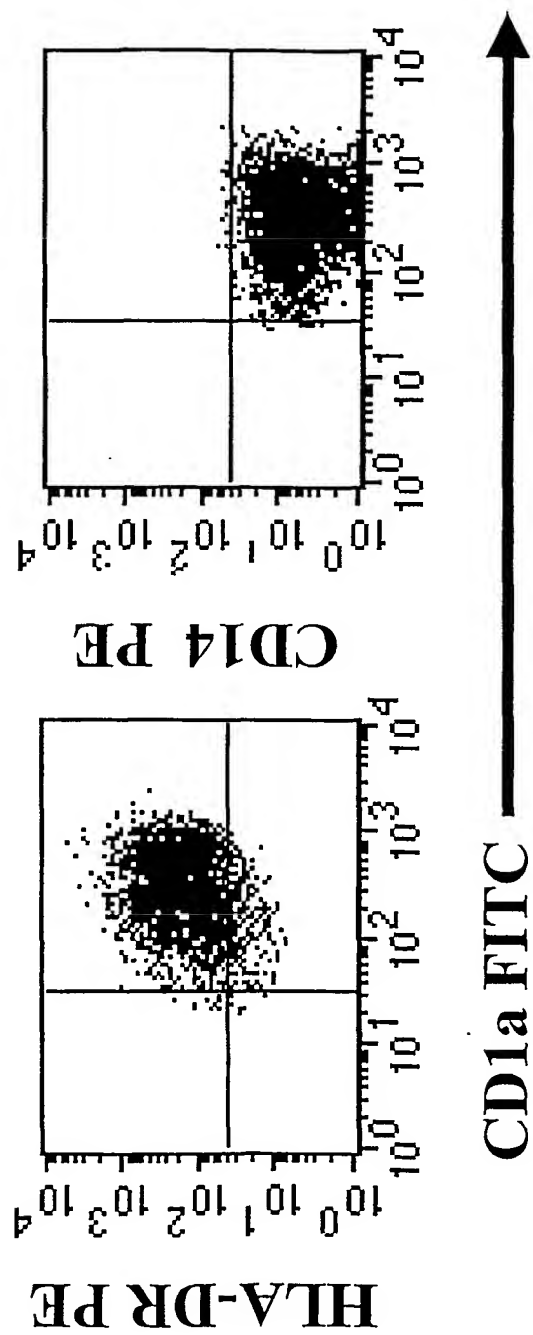
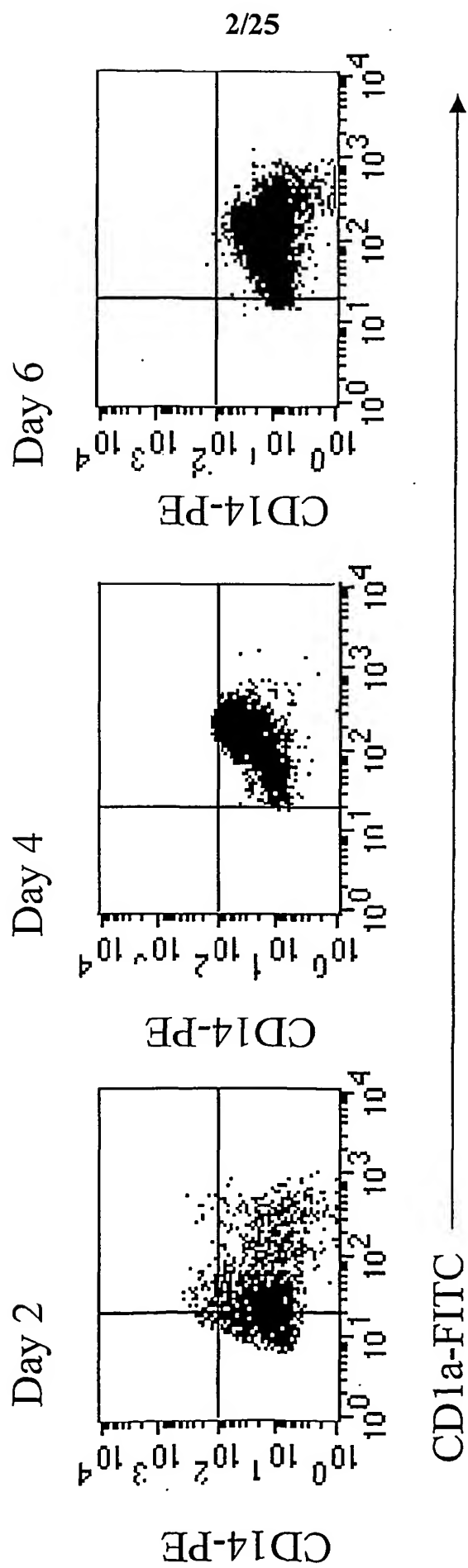
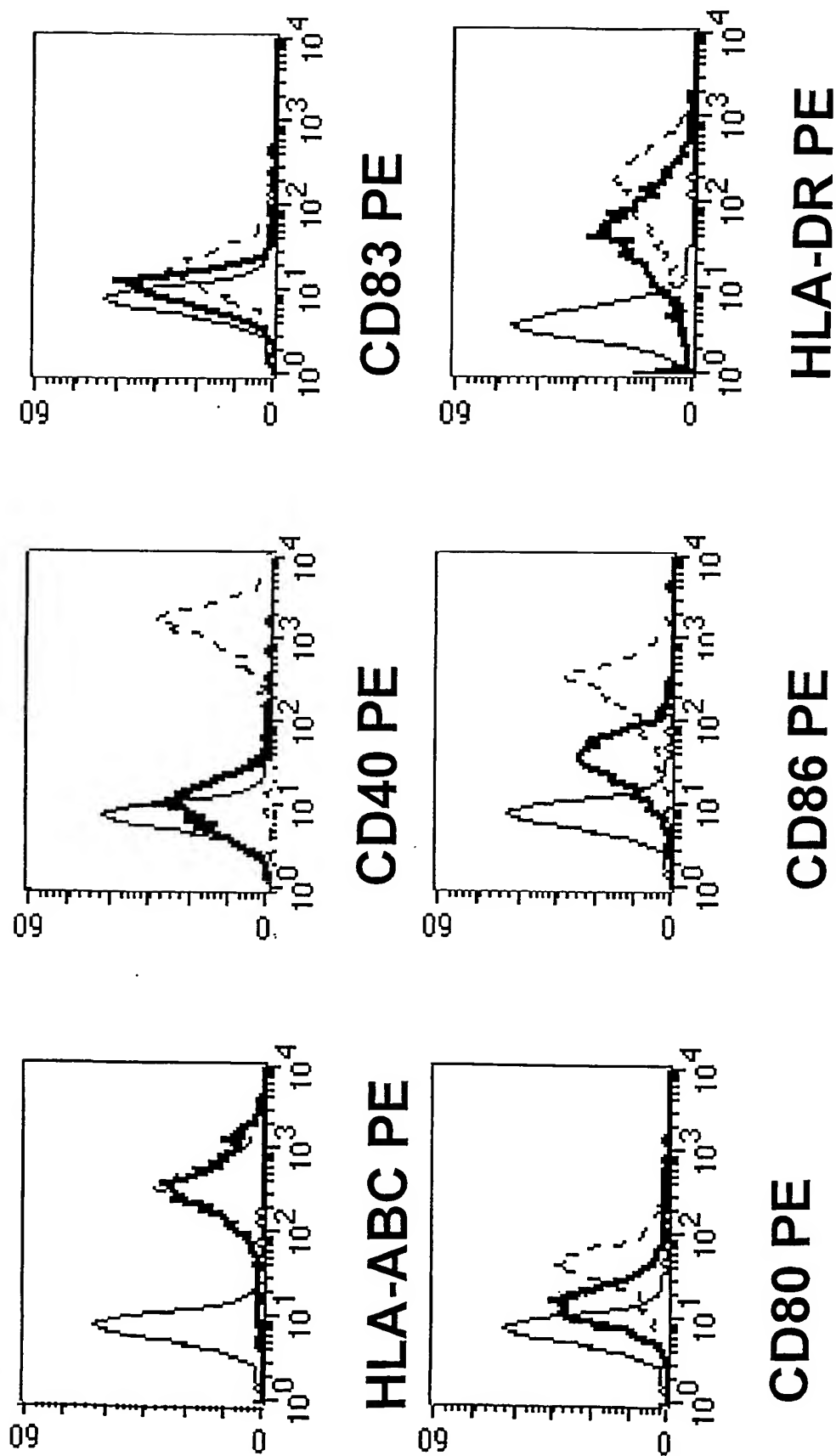


Figure 1B

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Figure 2A



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Figure 2B

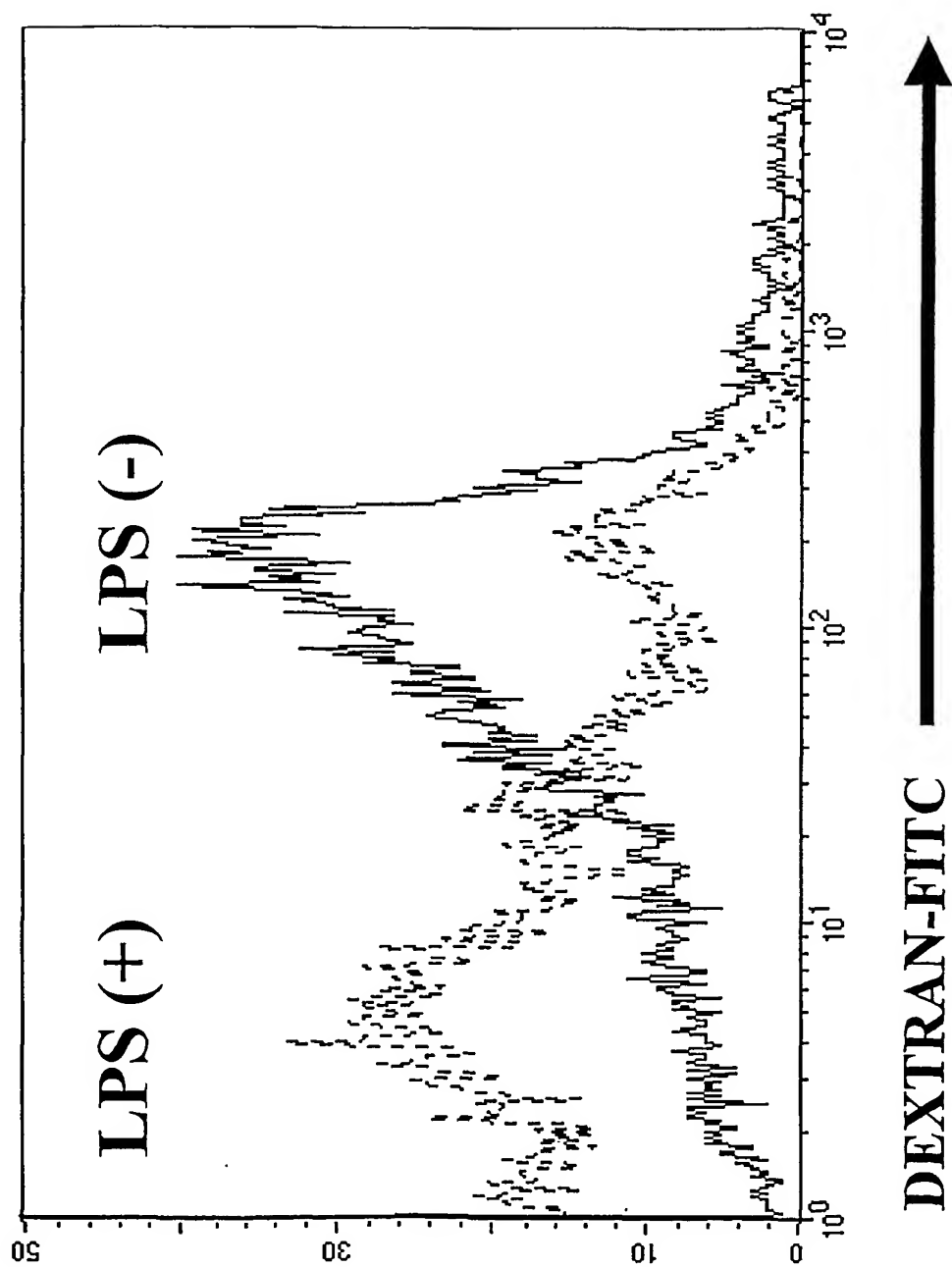
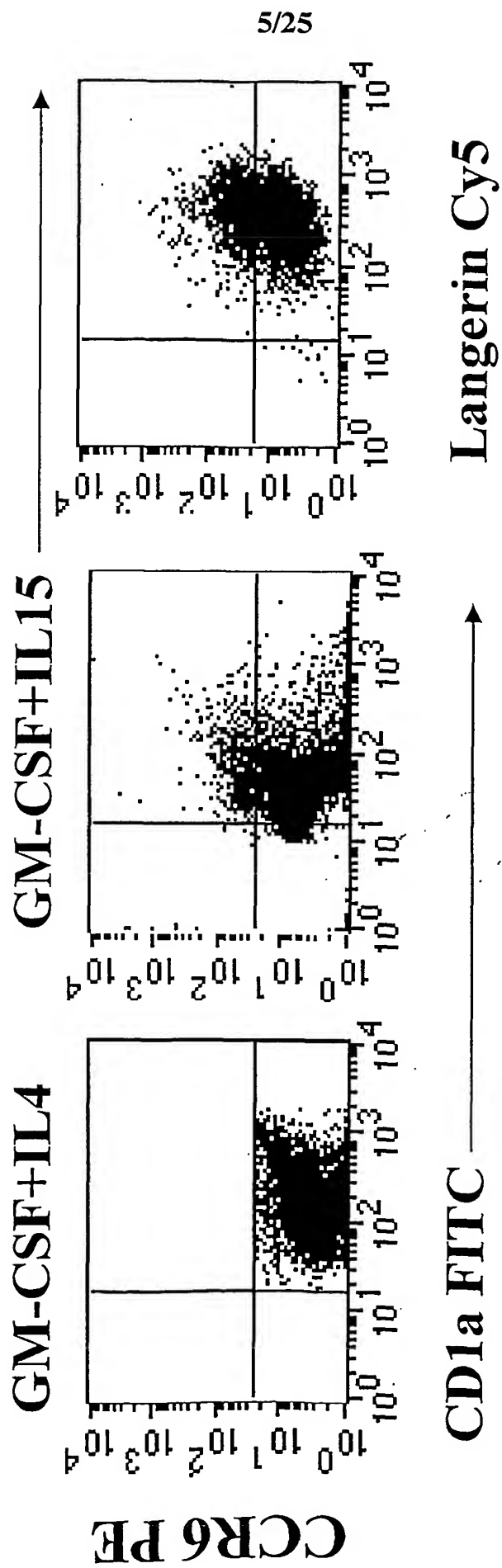
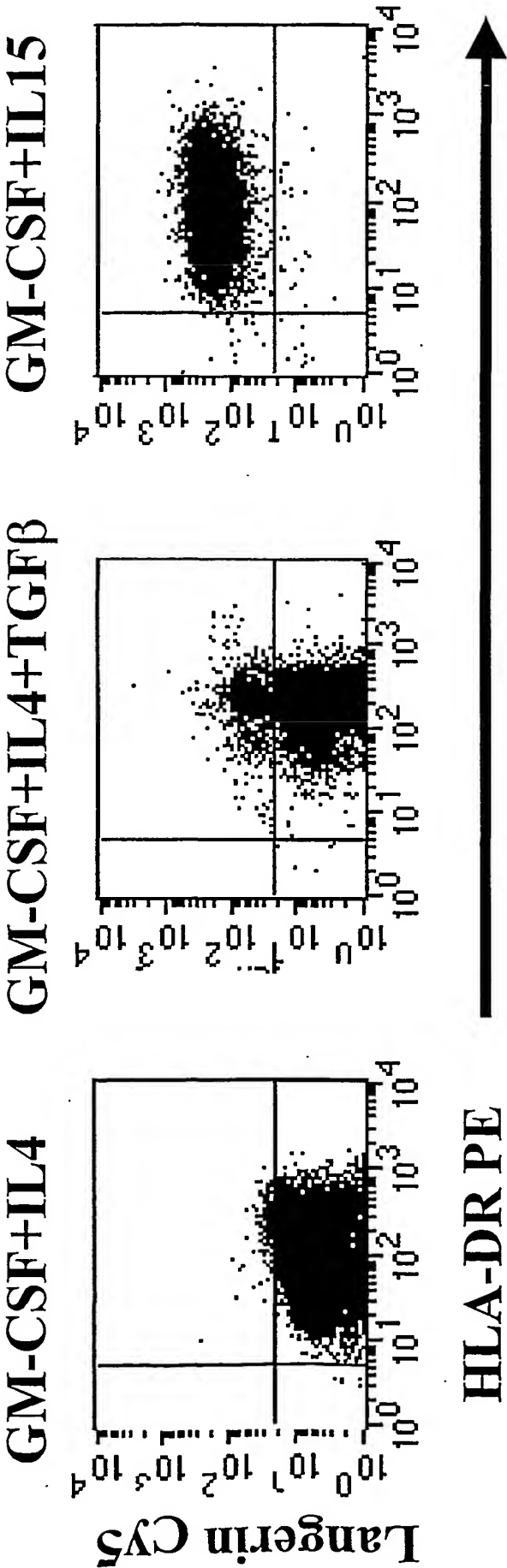


Figure 3A



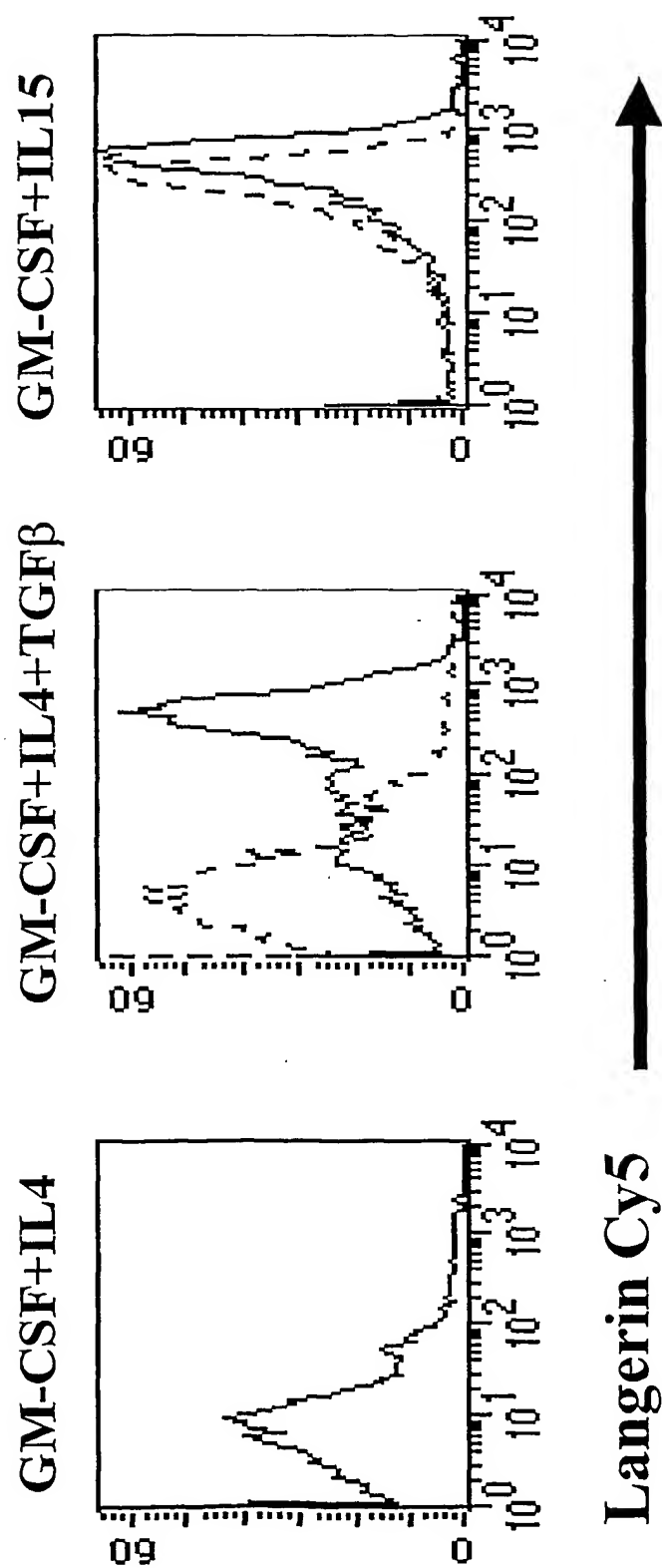
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Figure 3B



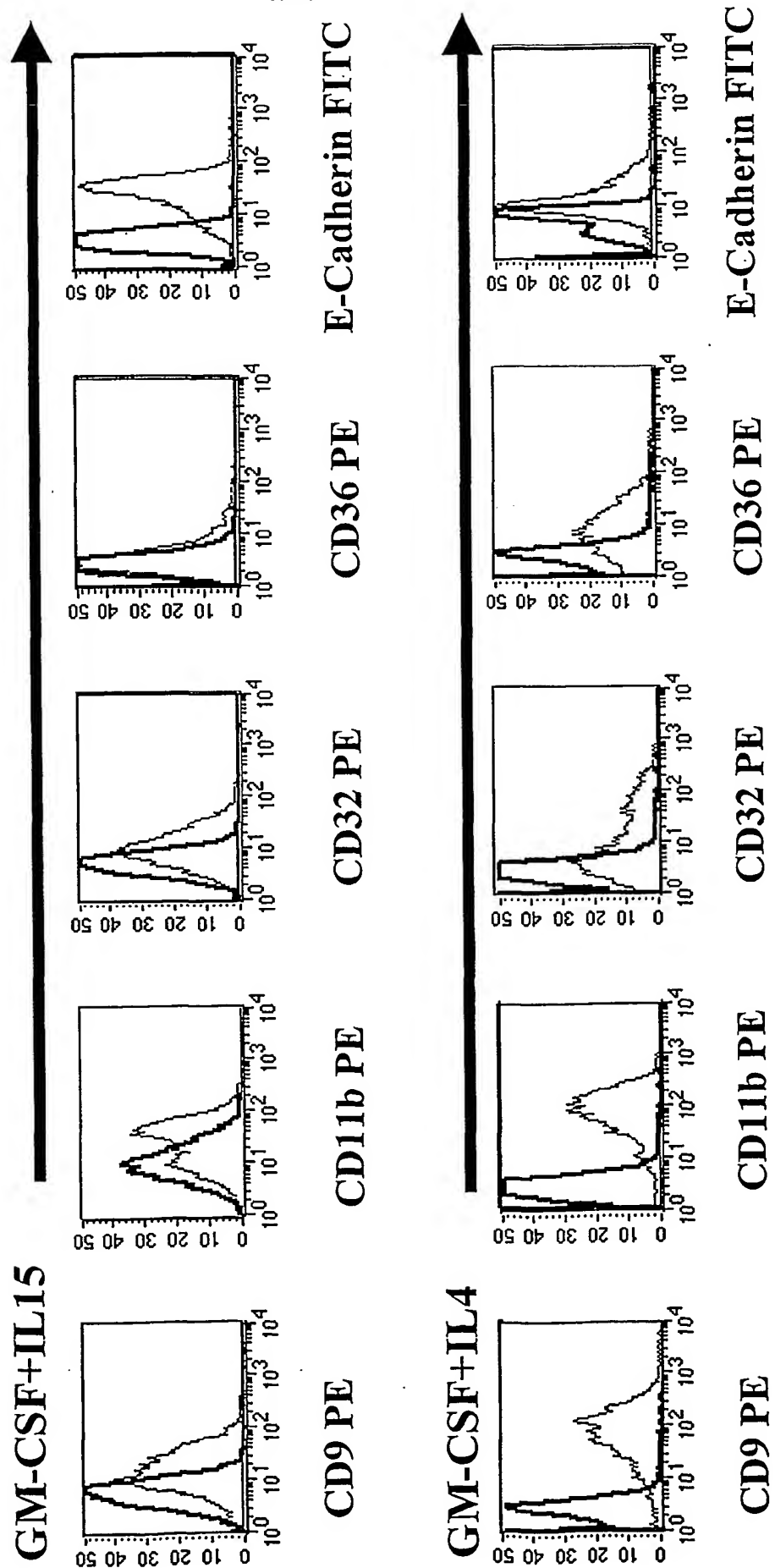
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Figure 3C

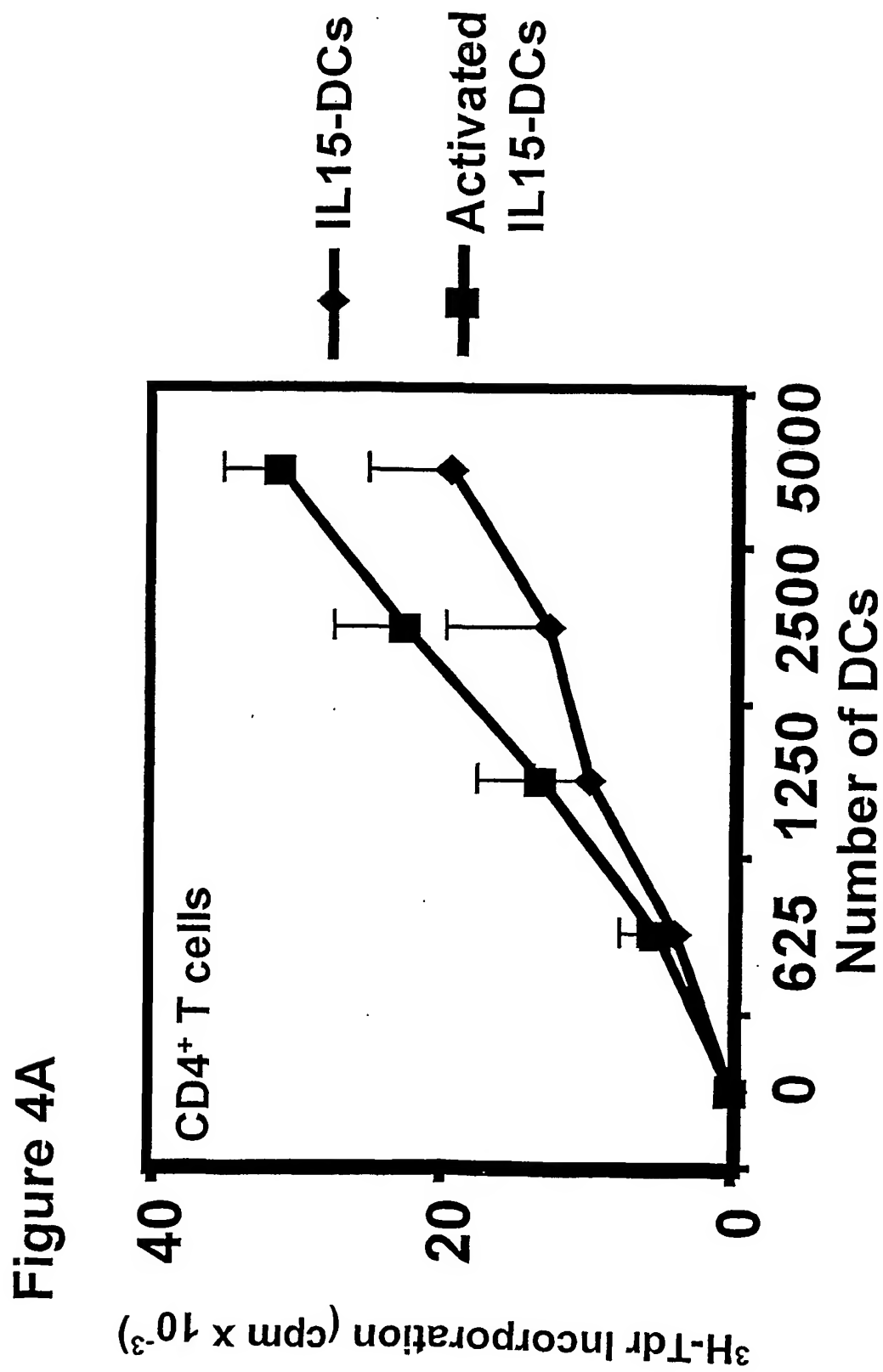


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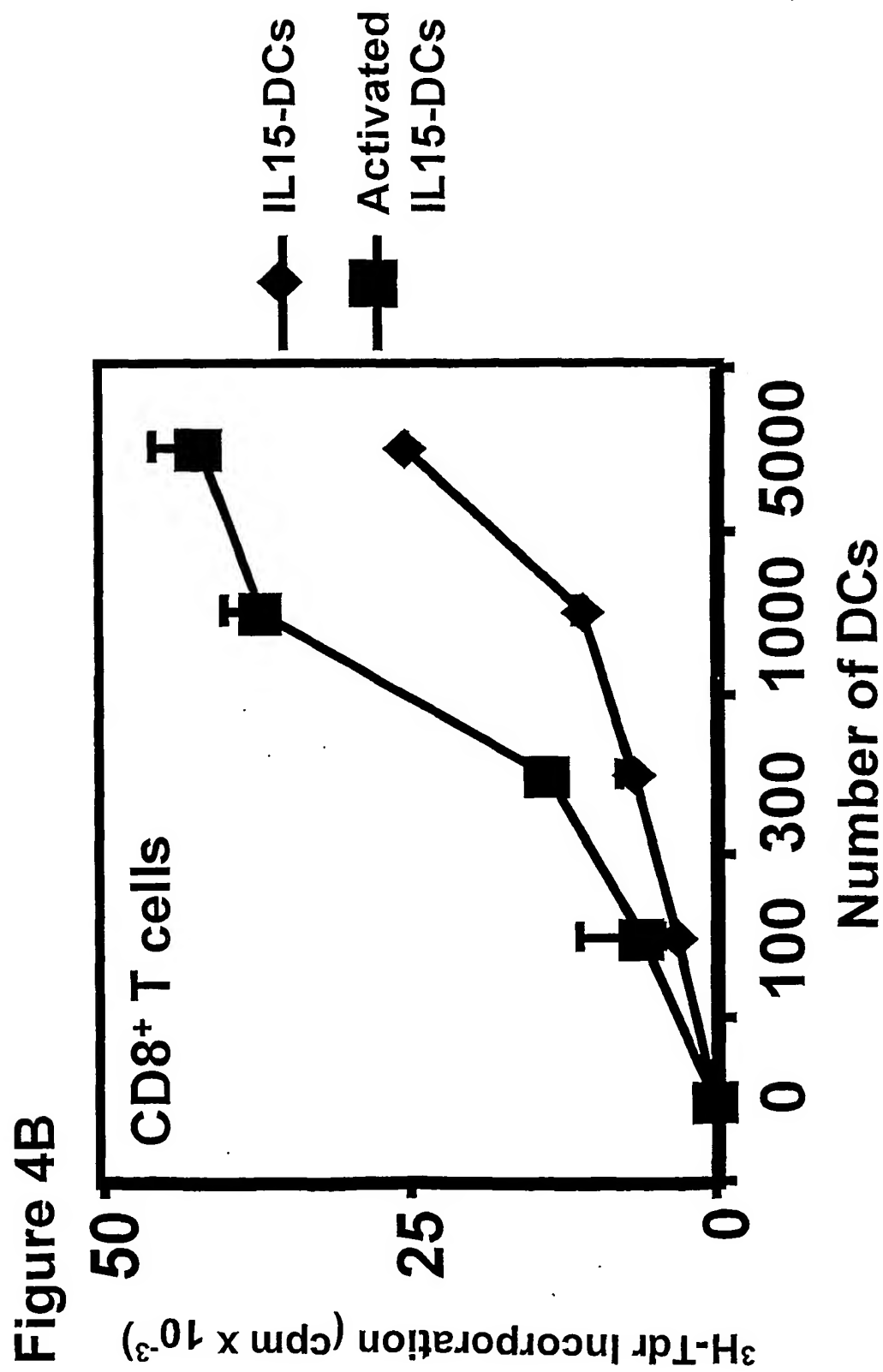
Figure 3D



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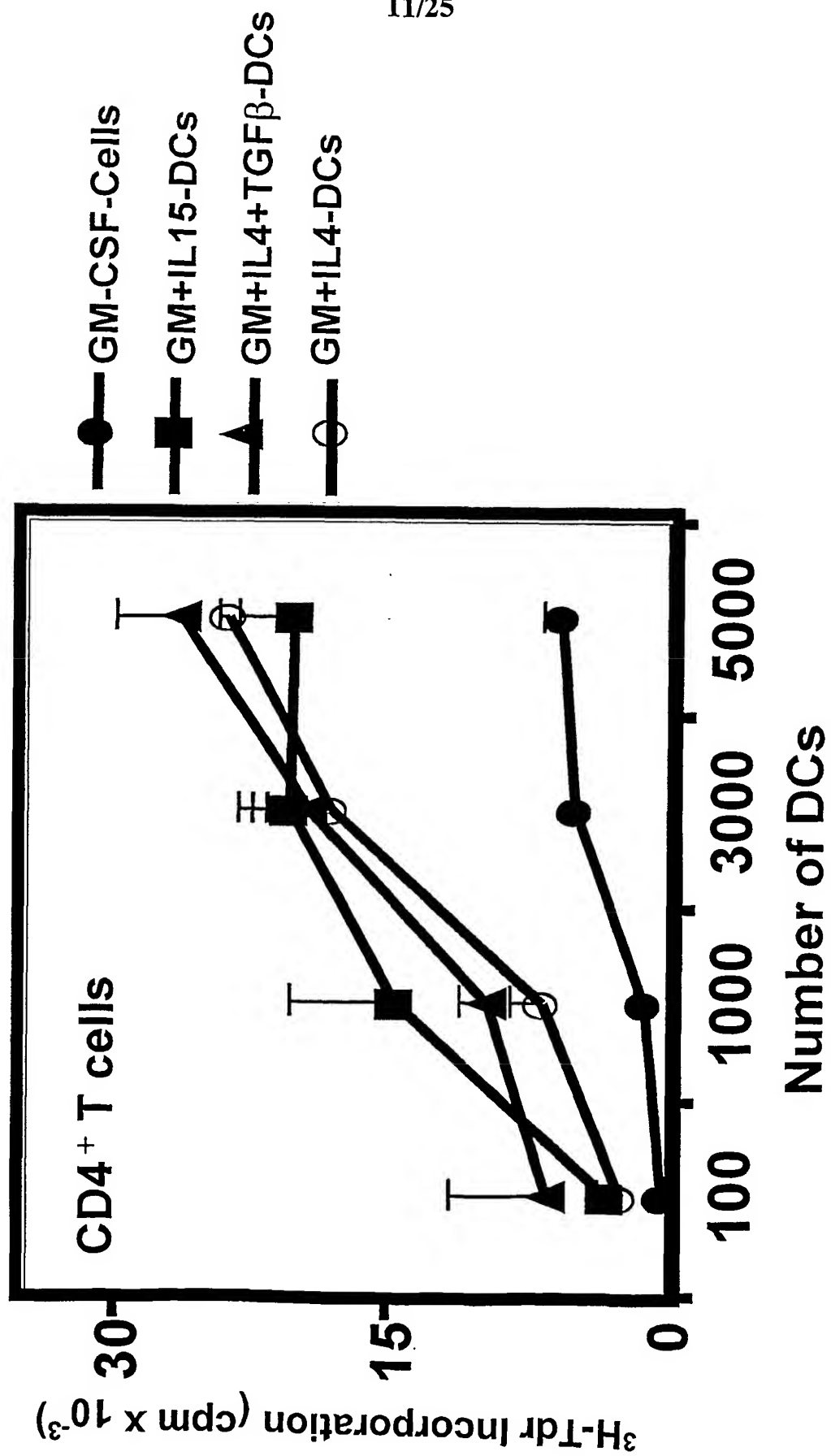


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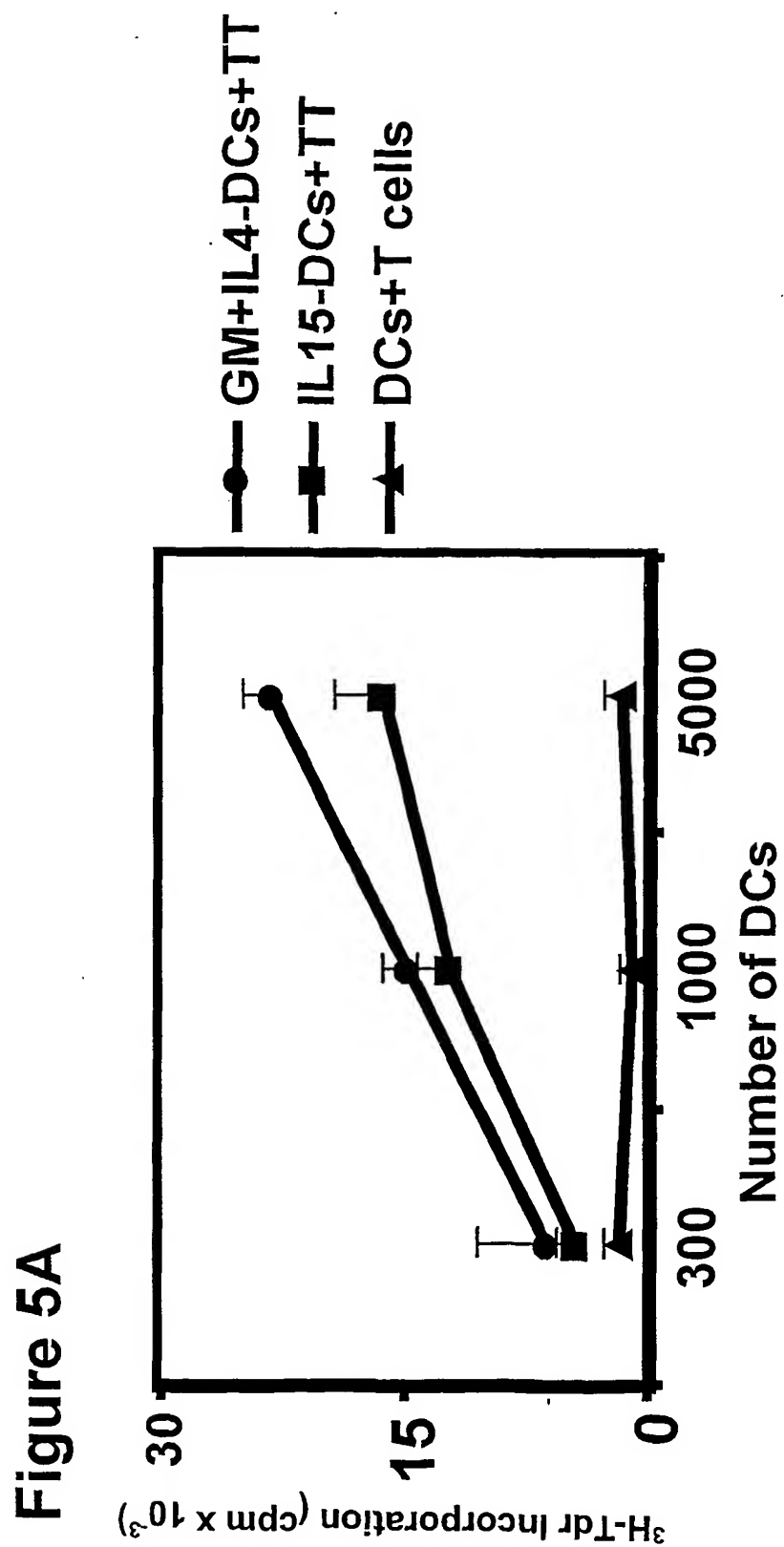


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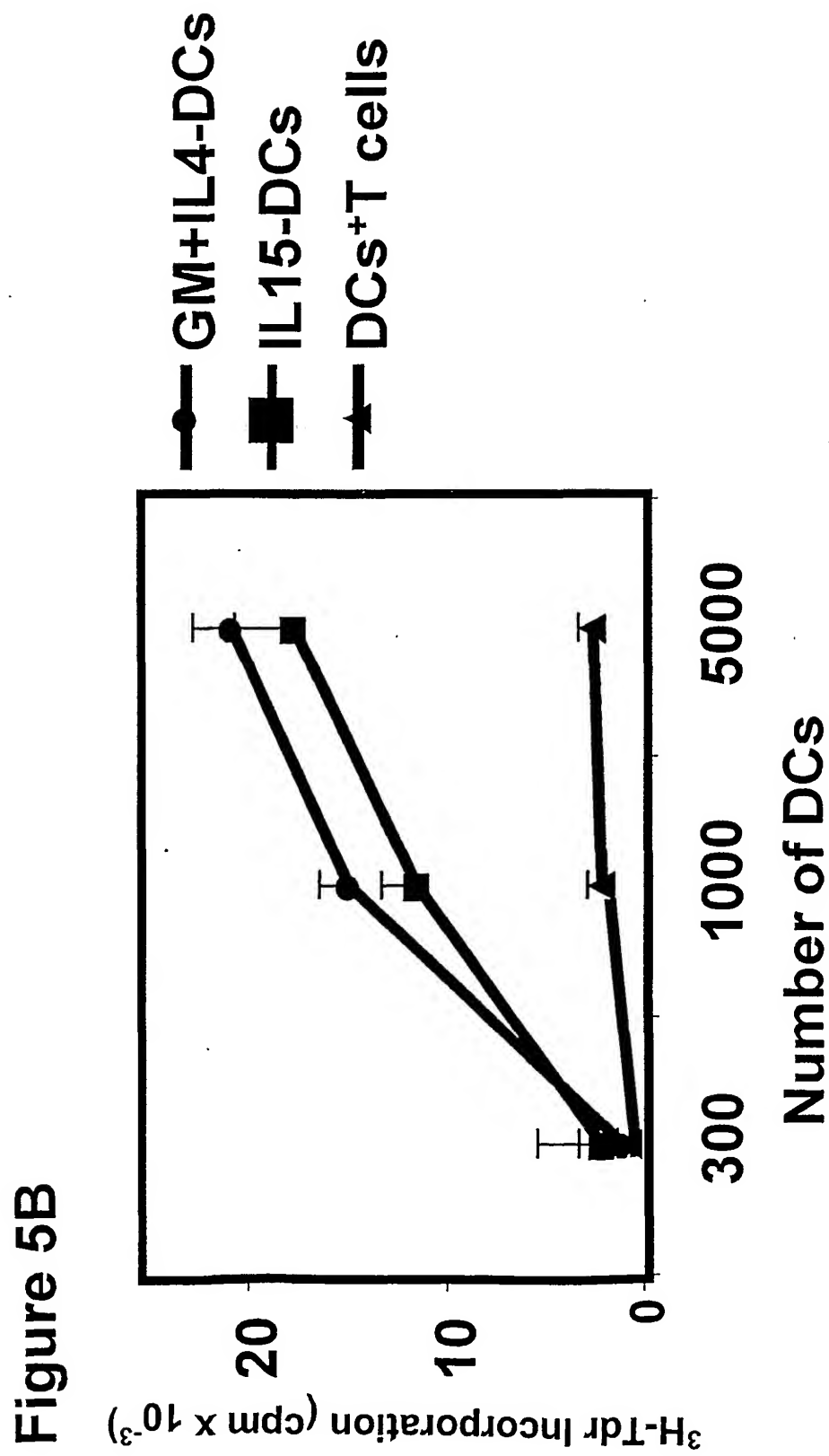
Figure 4C



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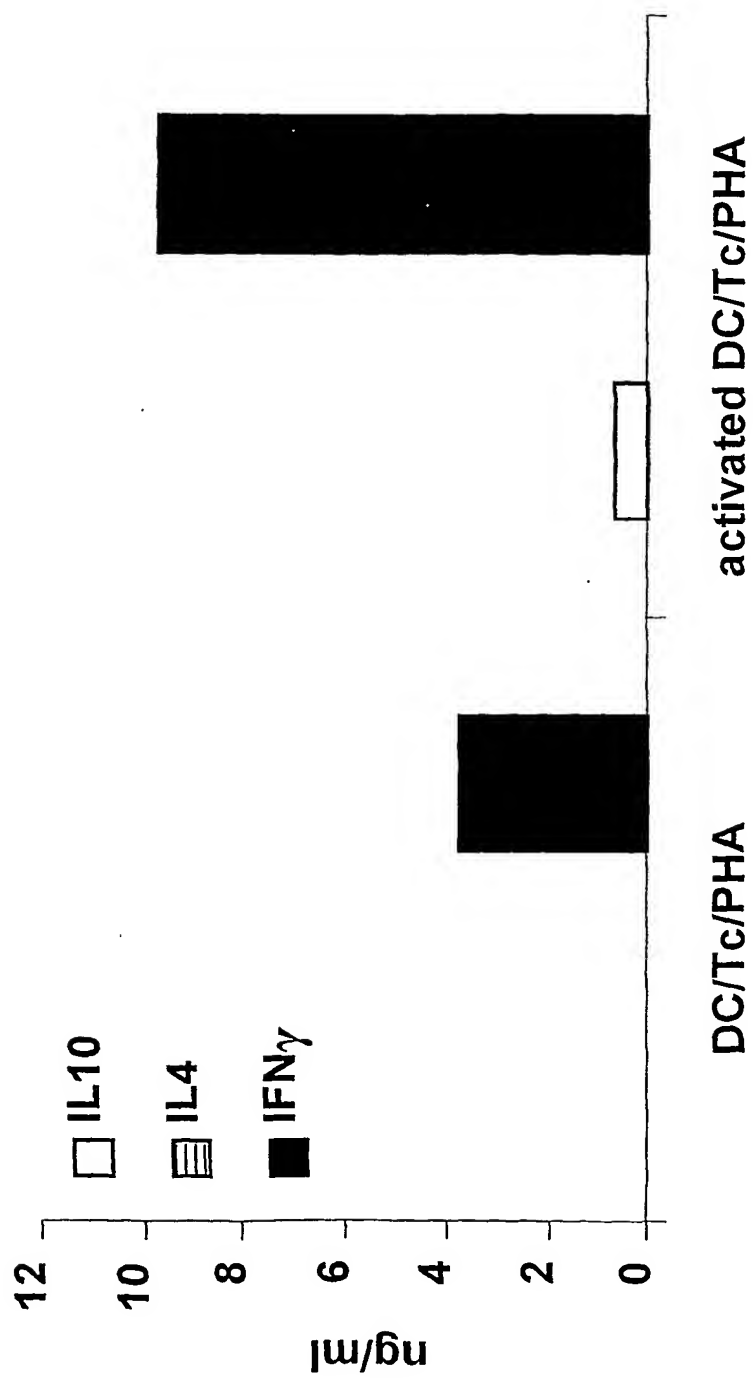


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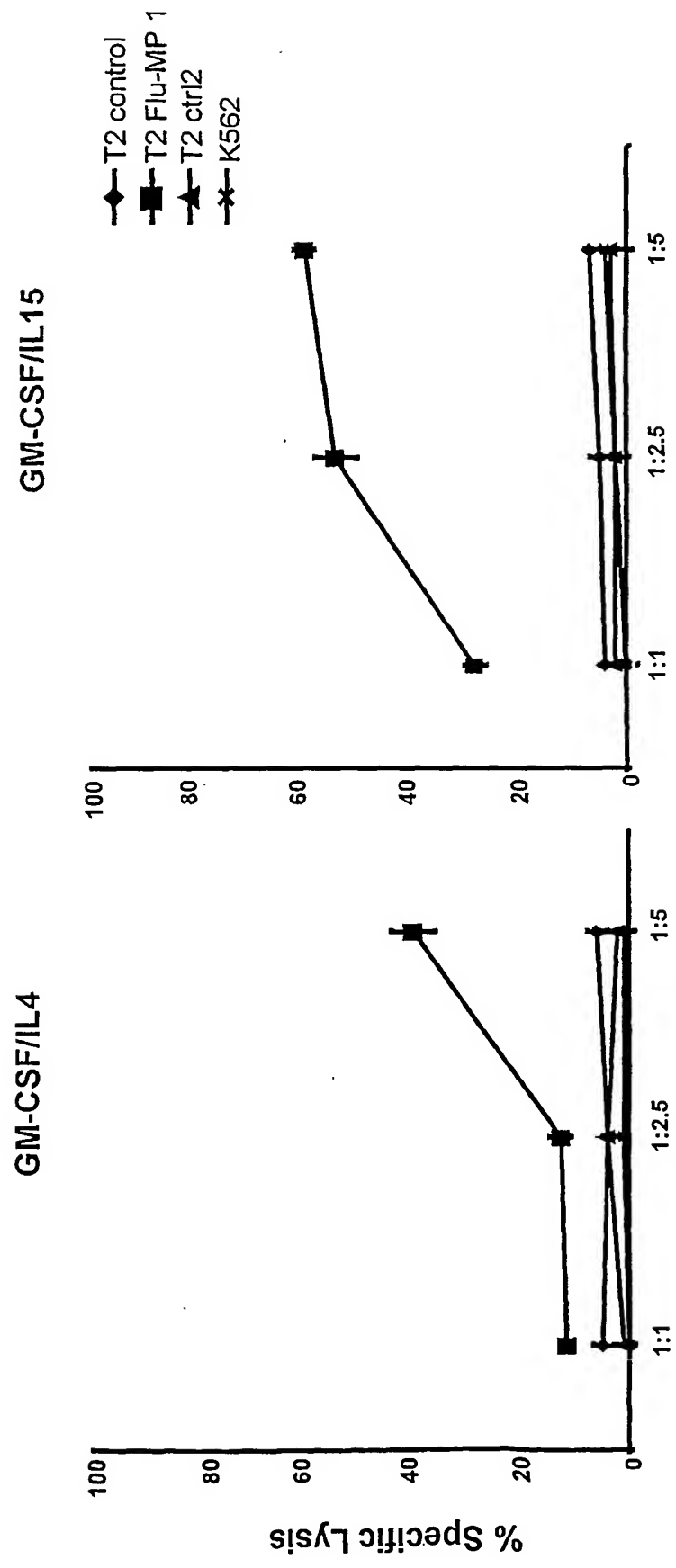
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Figure 6



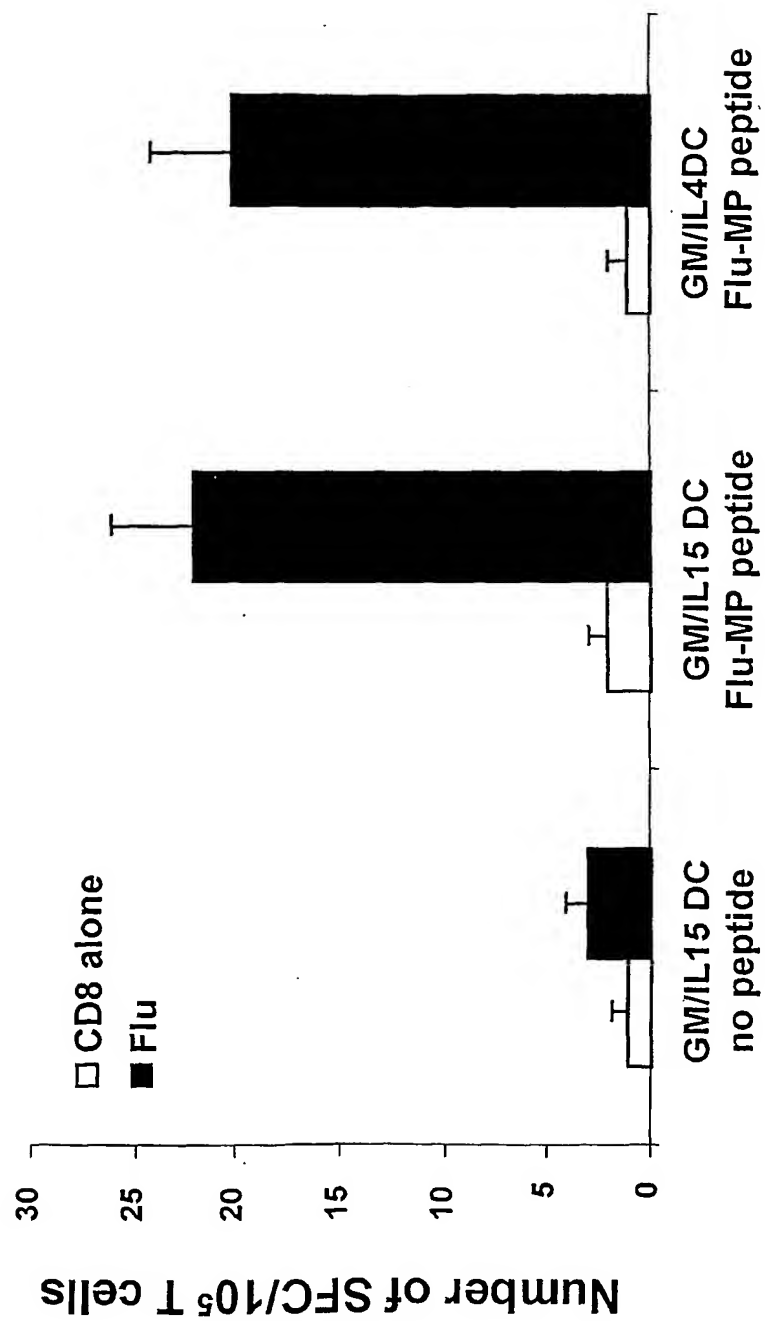
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Figure 7A



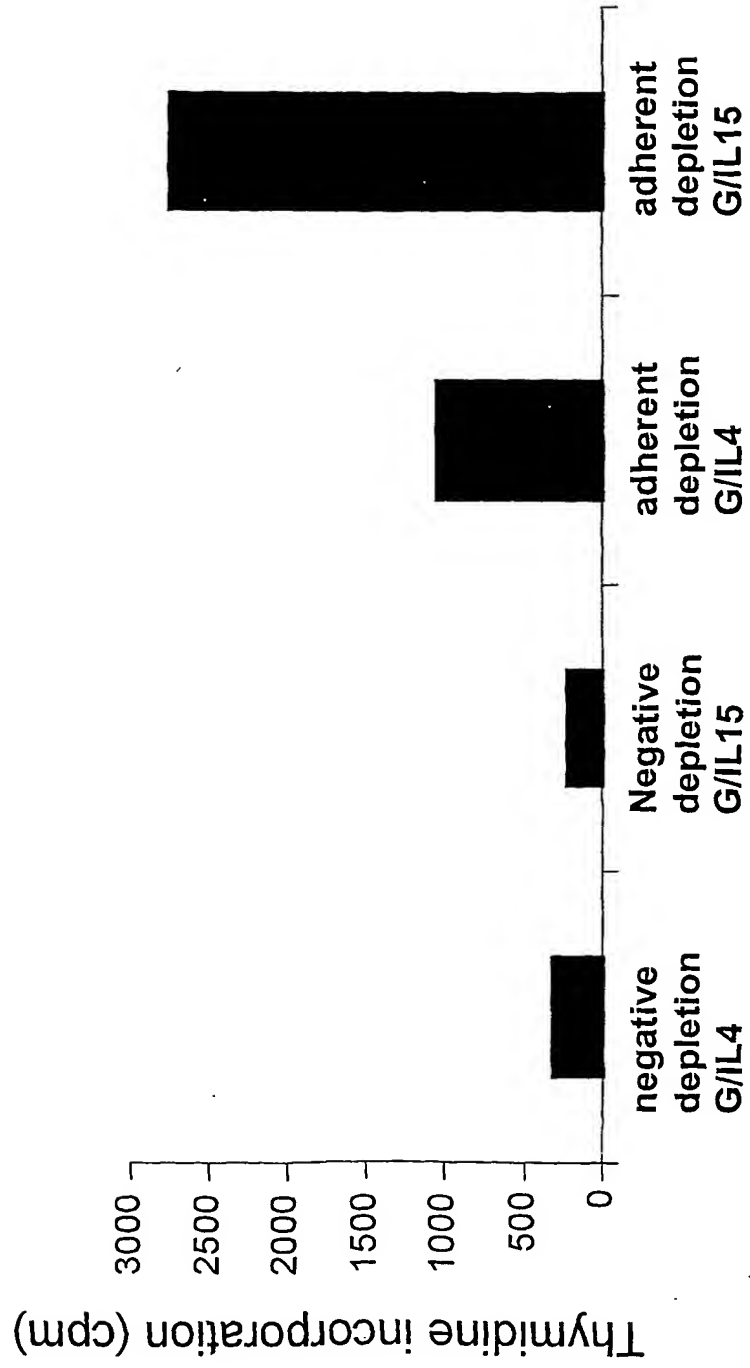
16/25

Figure 7B



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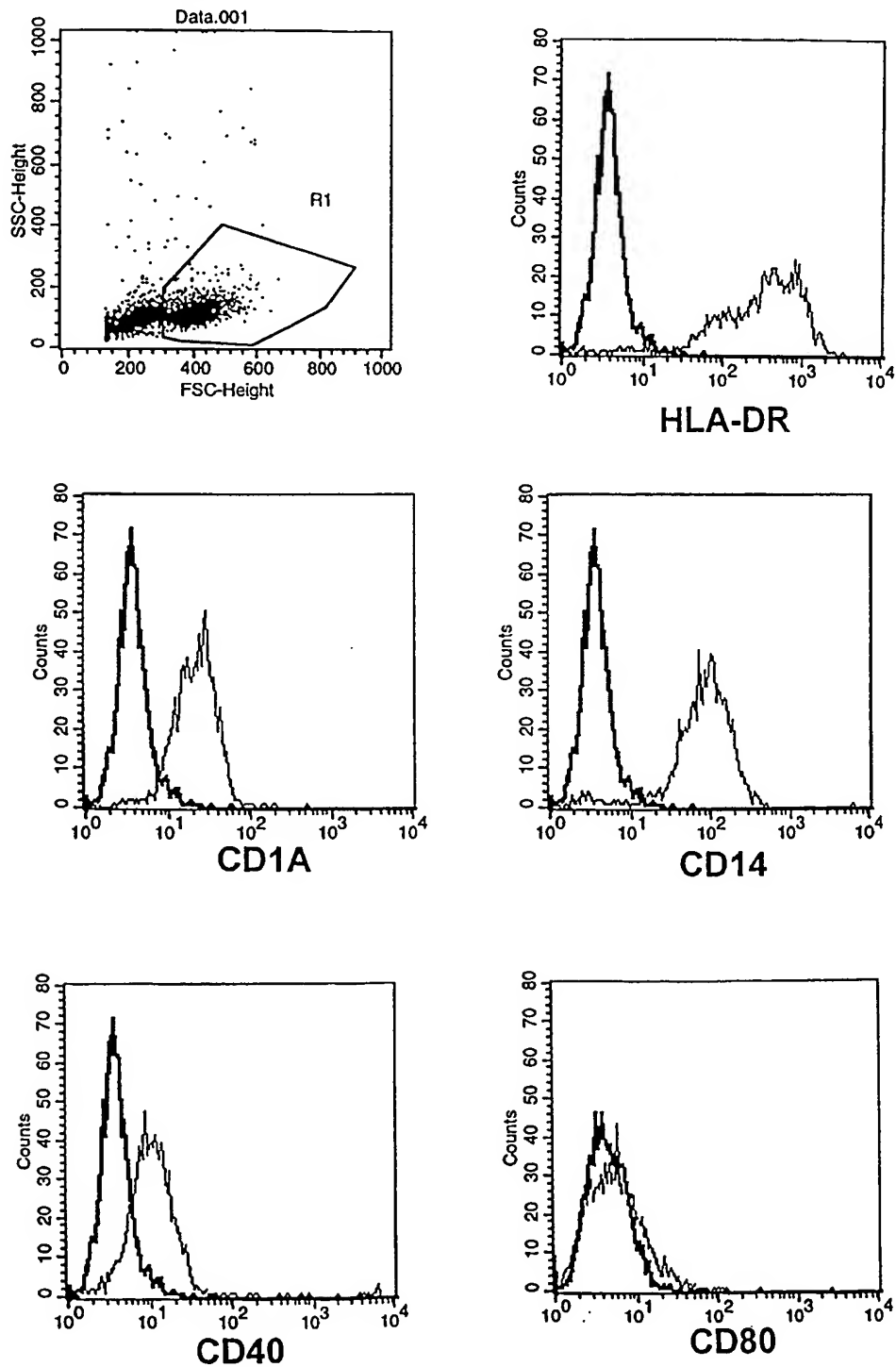
Figure 8



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Figure 9A

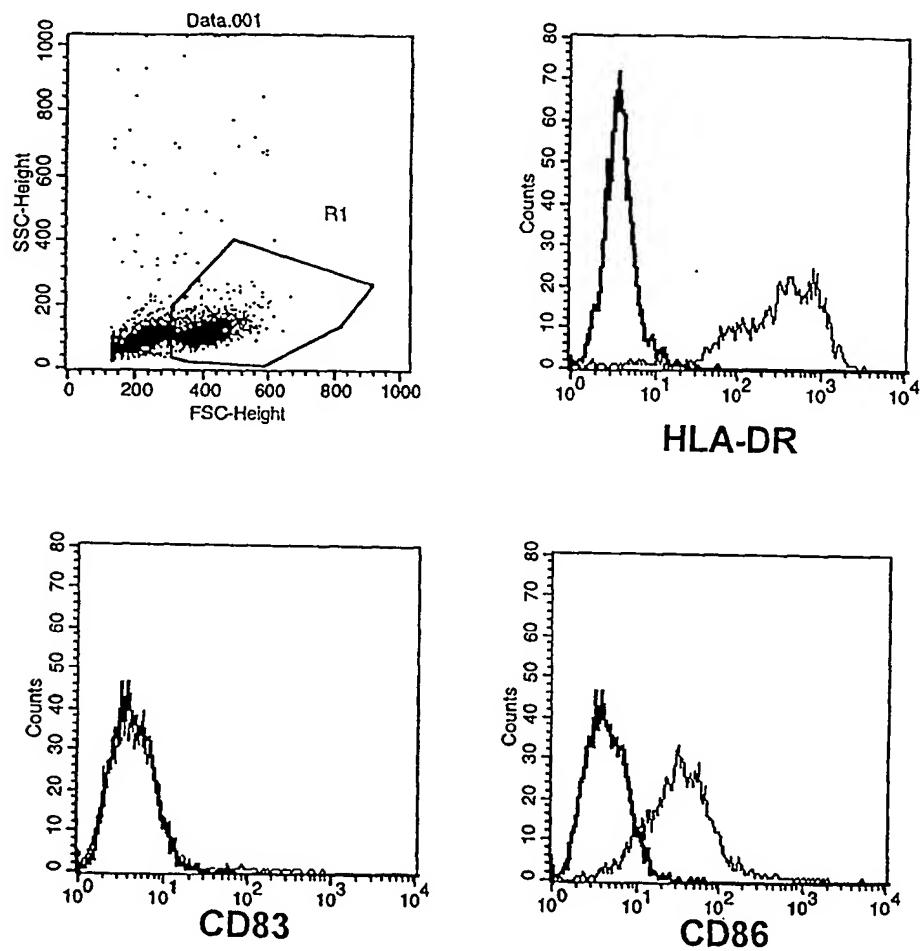
MONOCYTES CULTURED WITH IL-15 ALONE - DAY 2



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Figure 9B

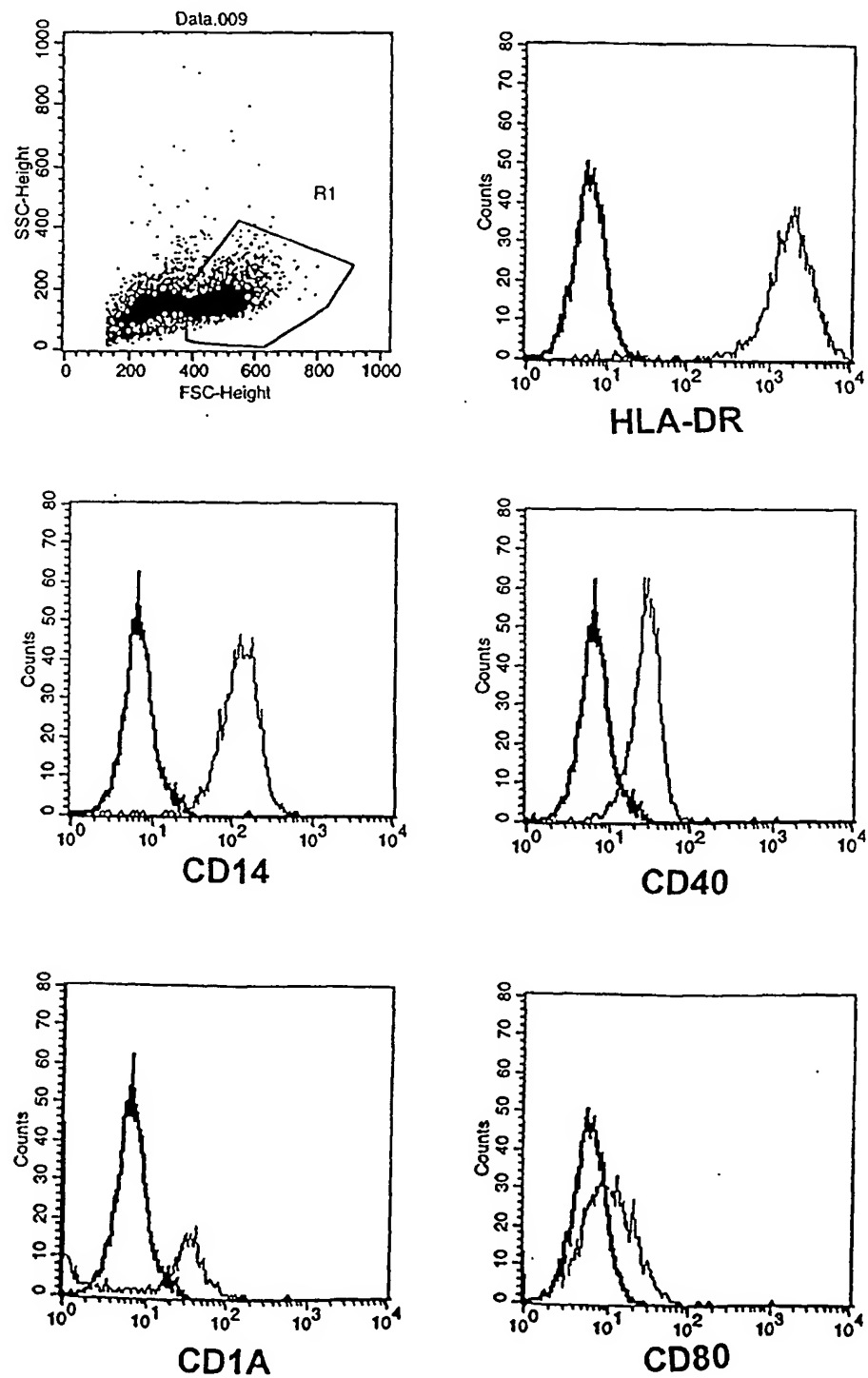
MONOCYTES CULTURED WITH IL-15 ALONE - DAY 2



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Figure 9C

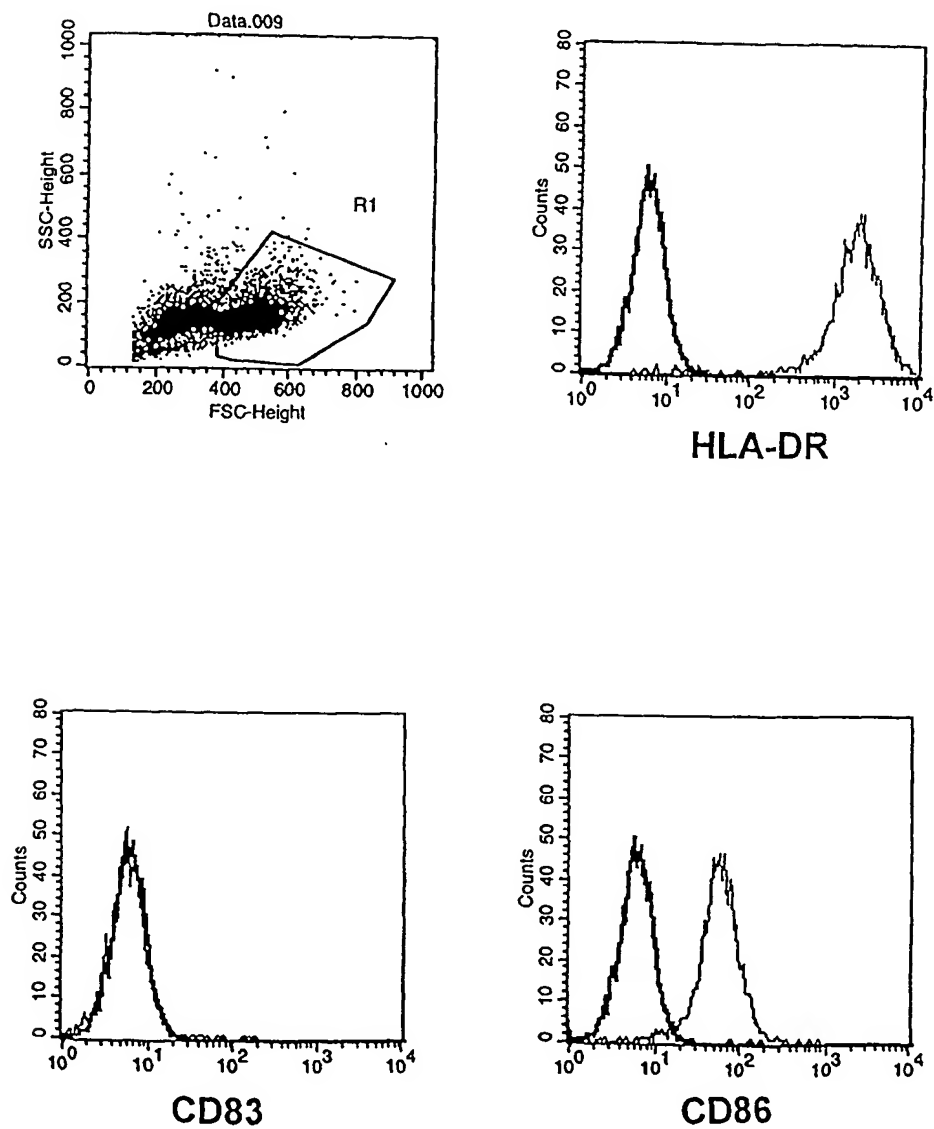
MONOCYTES CULTURED WITH IL-15 ALONE - DAY 4



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Figure 9D

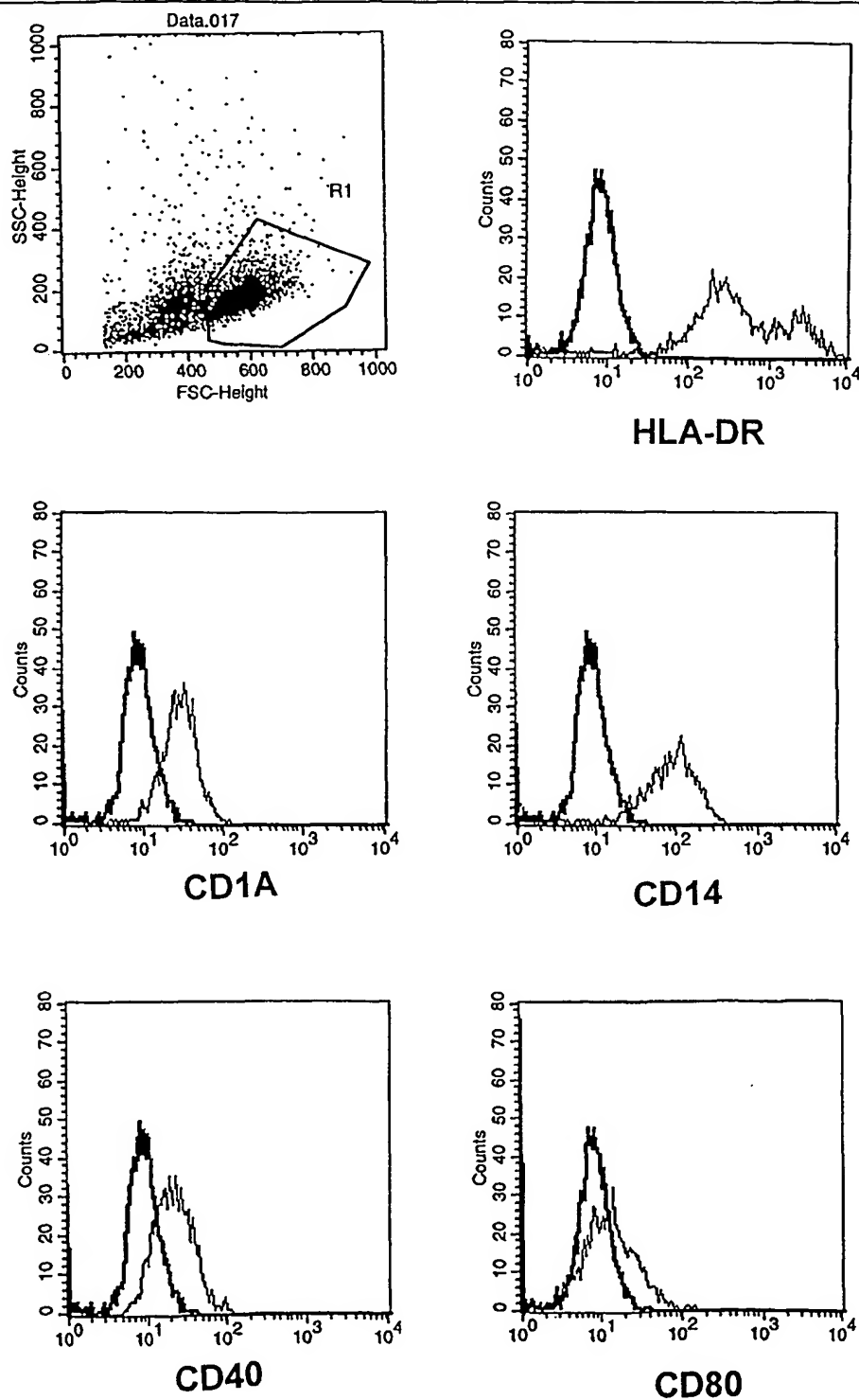
MONOCYTES CULTURED WITH IL-15 ALONE - DAY 4



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Figure 9E

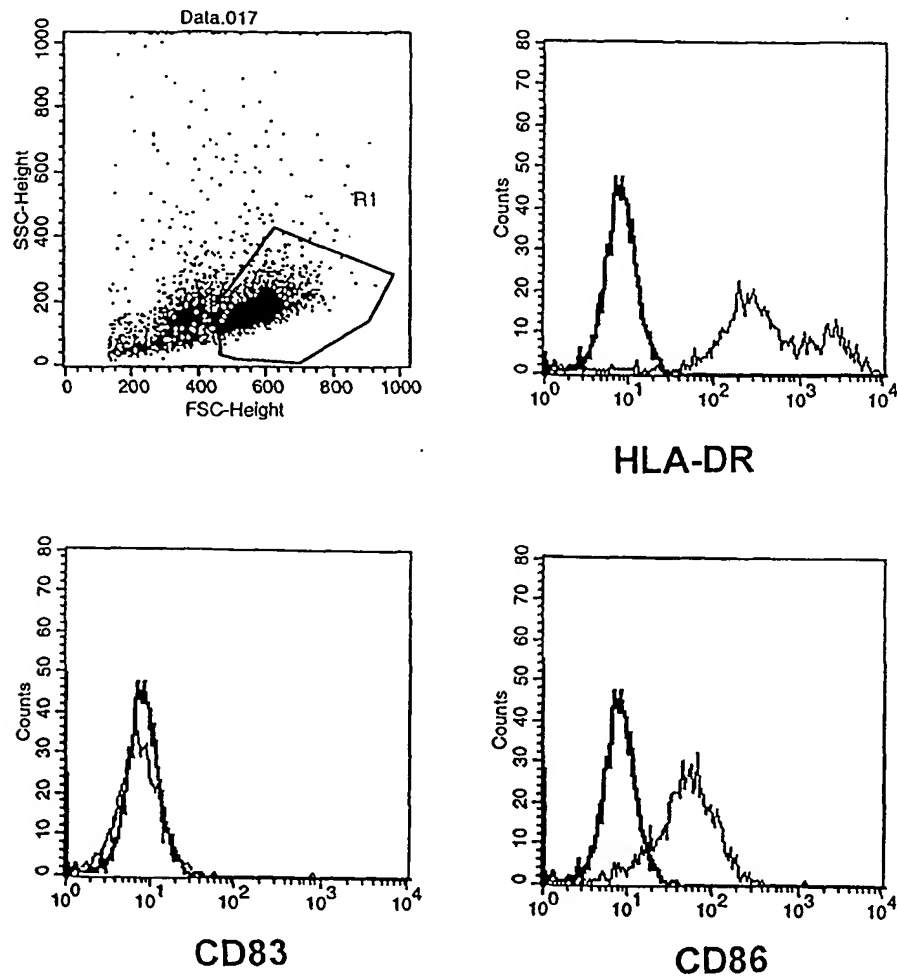
MONOCYTES CULTURED WITH IL-15 ALONE - DAY 6



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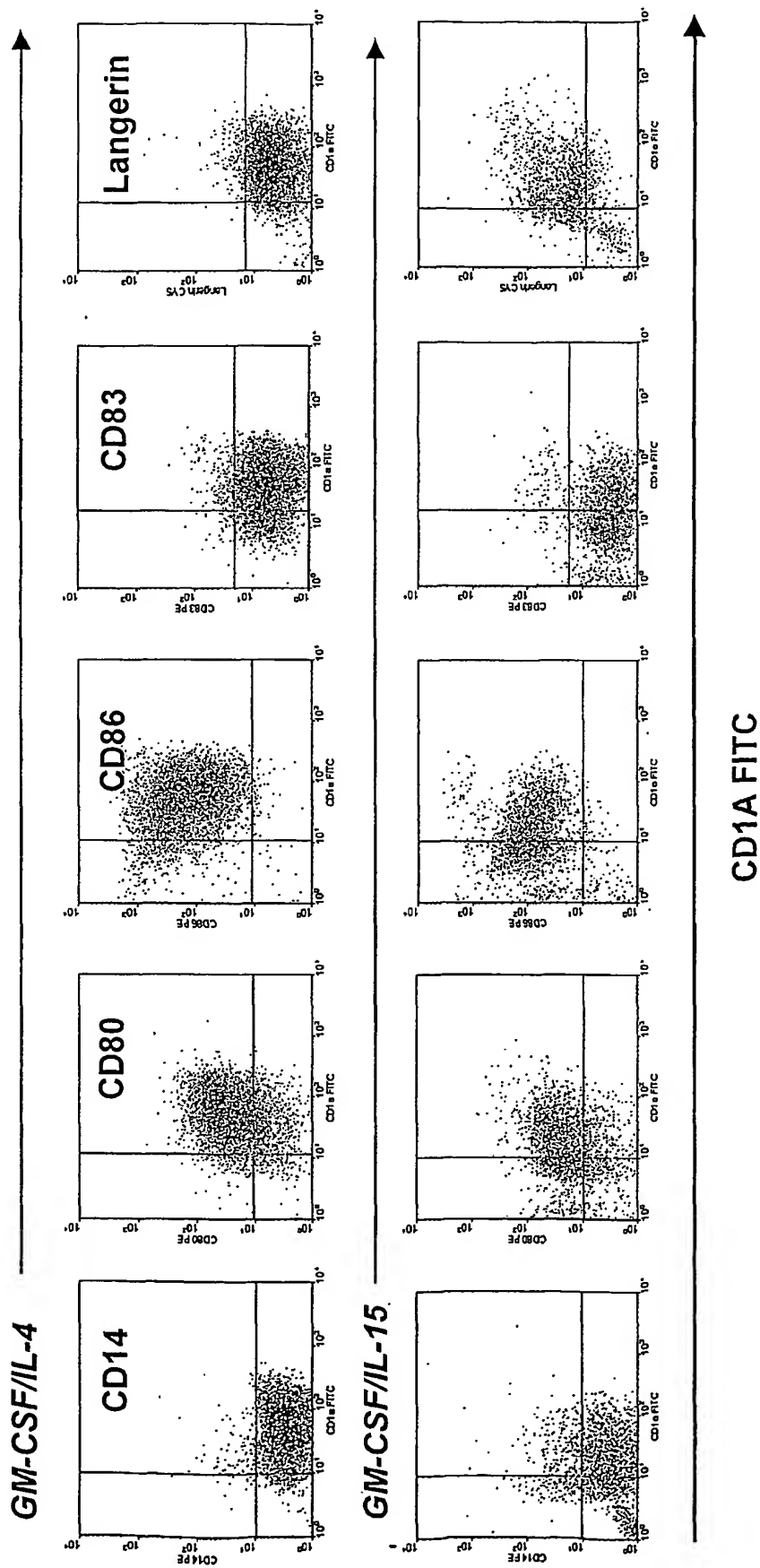
Figure 9F

MONOCYTES CULTURED WITH IL-15 ALONE - DAY 6



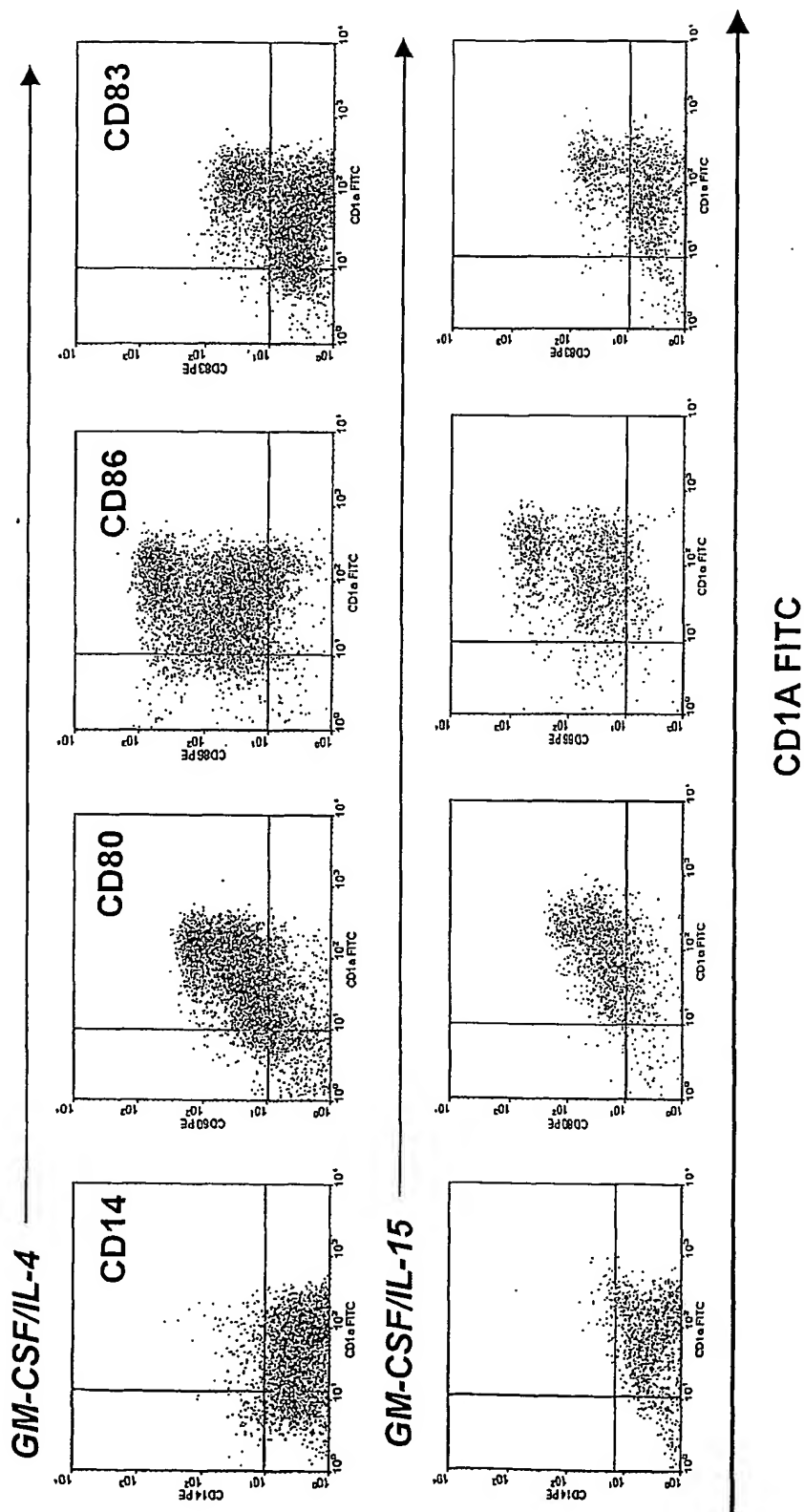
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Figure 10A



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Figure 10B



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